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(54) Title: PYRIMIDINE DERIVATIVES AND GUANINE DERIVATIVES, AND THEIR USE IN TREATING TUMOUR CELLS

R''XCHR'' (a)

(57) Abstract

The invention provides compounds exhibiting the ability to deplete Ob-alkylguanine-DNA alkyltransferase (ATase) activity in tumour cells. The compounds include certain pyrimidine derivatives of formula (II), wherein R is (i) a cyclic group having at least one 5- or 6-membered heterocyclic ring, optionally with a carbocyclic or heterocyclic ring fused thereto, the or each heterocyclic ring having at least one hetero atom chosen from O, N, or S, or a substituted derivative thereof; or (ii) phenyl or a substituted derivative thereof, R² is selected from H, C¹-C⁵ alkyl, halogen or NH², R⁴ and R⁵ which are the same or different are selected from H, NH-Y' or NOn wherein Y' is H, ribosyl, deoxyribosyl, arabinosyl, (a) wherein X is O or S, R" is alkyl and R''' is H or alkyl, or substituted derivatives thereof, n = 1 or 2 or R⁴ and R⁵ together with the pyrimidine ring form a 5-or 6-membered ring structure containing one or more hetero atoms, and pharmaceutically acceptable salts thereof. They include certain guanine derivatives of formula (XIII), wherein R⁶ is as defined at (i) for R above and Y' is as defined above.

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PYRIMIDINE DERIVATIVES AND GUANINE DERIVATIVES, AND THEIR USE IN TREATING TUMOUR CELLS

15 Technical Field

The present invention relates to pyrimidine derivatives and guanine derivatives, and their use in treating tumour cells. In particular, it relates to 6-hetarylalkyloxy pyrimidine derivatives, $\underline{0}^6$ -substituted guanine derivatives and \underline{S}^6 -substituted thioguanine derivatives, these compounds exhibiting the ability to deplete $\underline{0}^6$ -alkylguanine-DNA alkyltransferase (ATase) activity in tumour cells.

25 Background Art

It has been suggested to use $\underline{0}^6$ -alkyl guanine derivatives possessing $\underline{0}^6$ -alkylguanine-DNA alkyltransferase depleting activity in order to enhance the effectiveness of chemotherapeutic alkylating agents, principally those that methylate or chloroethylate DNA, used for killing tumour cells. There is increasing evidence that in mammalian cells the toxic and mutagenic effects of alkylating agents are to a large extent a consequence of alkylation at the $\underline{0}^6$ -position of guanine in DNA. The repair of $\underline{0}^6$ -alkylguanine is mediated by ATase, a repair protein that acts on the $\underline{0}^6$ -alkylated guanine residues by stoichiometric transfer of the alkyl group to a cysteine residue at the active site of the repair protein in an

autoinactivating process. The importance of ATase in protecting cells against the biological effects of alkylating agents has been most clearly demonstrated by the transfer and expression of cloned ATase genes or cDNAs into ATase deficient cells: this confers resistance to a variety of agents, principally those that methylate or chloroethylate DNA. Whilst details of the mechanism of cell killing by $\underline{0}^6$ -methylguanine in ATase deficient cells is not yet clear, killing by $\underline{0}^6$ -chloroethylguanine occurs through DNA interstrand crosslink formation to a cytosine residue on the opposite strand via a cyclic ethanoguanine intermediate, a process that is prevented by ATase-mediated chloroethyl group removal or complex formation.

The use of 0^6 -methylguanine and 0^6 -n-butylguanine for depleting ATase activity has been investigated (Dolan et al., Cancer Res., (1986) 46, pp. 4500; Dolan et al., Cancer Chemother.

Pharmacol., (1989) 25. pp 103. 0^6 -benzylguanine derivatives have been proposed for depleting ATase activity in order to render ATase expressing cells more susceptible to the cytotoxic effects of chloroethylating agents (Moschel et al., J. Med.Chem., 1992, 35, 4486). U.S. Patent 5 091 430 and International Patent Application No. WO 91/13898 Moschel et al. disclose a method for depleting levels of 0^6 -alkylguanine-DNA alkyl-transferase in tumour cells in a host which comprises administering to the host an effective amount of a composition containing 0^6 -benzylated guanine derivatives of the following formula:

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and R^a is a benzyl group or a substituted benzyl group. A benzyl group may be substituted at the ortho, meta or para position with a substituent group such as halogen, nitro, aryl such as phenyl or substituted phenyl, alkyl of 1-4 carbon atoms, alkoxy of 1-4 carbon atoms, alkenyl of up to 4 carbon atoms, alkynyl of up to 4 carbon atoms, amino, monoalkylamino, dialkylamino, trifluoromethyl, hydroxy, hydroxymethyl, and SO_nR^b wherein n is 0, 1, 2 or 3 and R^b is hydrogen, alkyl of 1-4 carbon atoms or aryl. Chae <u>et al</u>., J.Med.Chem., 1994, 37, 342-347 describes tests on $\underline{0}^6$ -benzylguanine analogs bearing increasingly bulky substituent groups on the benzene ring or at position 9. Chae et. al., <u>J. Med.</u> <u>Chem.</u> **1995**, 38, 359-365 describe several 8-substituted $\underline{0}^6$ -benzylguanines, 2- and/or 8-substituted 6-(benzyloxy)purines, substituted 6(4)-(benzyloxy)pyrimidines, and a 6-(benzyloxy)-s-triazine which were tested for their ability to inactivate ATase. Two types of compounds were identified as being significantly more effective than Q^6 -benzylquanine at inactivating ATase in human HT29 colon tumour cell extracts. These were 8-substituted $\underline{0}^6$ -benzylguanines bearing electron-withdrawing groups at the 8-position (e.g. 8-aza-0⁶-benzylguanine and 0^6 -benzyl-8-bromoquanine) and 5-substituted 2,4-diamino-6-(benzyloxy)pyrimidines bearing electron withdrawing groups at the 5-position (e.g. 2,4-diamino -6-(benzyloxy)-5-nitrosoand 2,4-diamino-6-(benzyloxy)-5-nitropyrimidine). The latter derivatives were also more effective than Q^{b} -benzylguanine at inactivating ATase in intact HT29 colon tumour cells. WO 96/04280 published after the priority dates of this application concerns similar substituted 0^6 -benzylguanines and 30 6(4)-benzyloxypyrimidines.

The present Applicants are also Applicants in International Patent Application PCT/IE94/00031 which was published under No. WO 94/29312. WO 94/29312 (the contents of which are incorporated herein by reference in their entirety) describes $\underline{0}^6$ -substituted guanine derivatives of formula I:

wherein

Y is H, ribosyl, deoxyribosyl, or R''XCHR''', wherein X is O or S, R'' and R''' are alkyl, or substituted derivatives thereof;

R' is H, alkyl or hydroxyalkyl;

R is (i) a cyclic group having at least one 5- or 6-membered heterocyclic ring, optionally with a carbocyclic or heterocyclic ring fused thereto, the or each heterocyclic ring having at least one hetero atom chosen from O, N, or S, or a substituted derivative thereof; or

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(ii) naphthyl or a substituted derivative thereof;

and pharmaceutically acceptable salts thereof.

- In order to be useful for depleting ATase activity and thus enhance the effects of the above-mentioned chemotherapeutic agents, compounds should have combination of characteristics assessed by reference to:
- 30 1) In vitro inactivation of recombinant ATases.
 - 2) Stability.
 - 3) Solubility.

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4) Inactivation of ATase in mammalian cells and/or tumour xenografts.

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5) Sensitization of mammalian cells and/or tumour xenografts to the killing or growth inhibitory effects of the said chemotherapeutic agents

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The behaviour of novel compounds in this combination of tests is unpredictable. Molecular interactions including steric factors in the unpredictability of ATase inactivation may be related to the nature of the environment of the cysteine acceptor site in the ATase molecule.

The structure of the ATase protein derived from E. coli (Ada gene) has been elucidated by X-ray crystallographic techniques (M.H. Moore et. al., EMBO Journal, 1994, 13, 1495.). While the amino acid sequence of human ATase differs somewhat from that of 15 bacterial origin, all known ATases (human, rodent, yeast, bacterial) contain the cysteine (Cys) acceptor site in a common fragment, Pro-Cys-His-Arg. A homology model of human ATase generated by computer from the crystal structure of the Ada protein (J.E.A. Wibley et. al., Anti-Cancer Drug Design, 1995, 10, 75.) resembles it in having the Cys acceptor buried in a pocket deep in the protein. Considerable distortion of the structure is necessary to bring either an 0^6 -alkylated quanine residue in intact DNA, or even free quanine alkylated by a relatively large group like benzyl, close to the Cys acceptor. These configurational changes are initiated by a characteristic binding of duplex DNA to the protein (K. Goodtzova et. al. Biochemistry, 1994, 33, 8385).

Since the amino acid components and dimensions of the ATase active site "pocket" are still unknown as are the details of the mechanism involved, it is impossible to predict the activity of a particular $\underline{0}^6$ -alkylated guanine or analogous ring system.

Published work in this field relates predominantly to the use of $\underline{0}^6$ -alkyl guanine derivatives having a nucleus identical to that of guanine in DNA. Chae <u>et</u>. <u>al</u>., <u>J</u>. <u>Med</u>. <u>Chem</u>. 1995, 38, 359-365 have described tests on a limited number of compounds in which the

guanine ring was modified. However these compounds all had benzyl substitution at the $\underline{0}^6$ - position of the modified guanine ring or 6(4)-benzyloxy substitution on the pyrimidine ring. The observation that subtle changes in the substituents on the guanine ring or in the purine skeleton can generate agents that are very ineffective ATase inactivators, in comparison with their "parent" structure, suggests that more substantial modifications might also disrupt the ATase inactivating function.

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There is a need for additional novel compounds useful for depleting ATase activity in order to enhance the effects of chemotherapeutic agents such as chloroethylating or methylating anti-tumour agents. It is a further object to provide compounds having better ATase inactivating characteristics than 0^6-benzylguanine and having different solubility patterns.

Another object of the invention is to provide pharmaceutical compositions containing compounds which are useful for depleting ATase activity. A further object of the present invention is to provide a method for depleting ATase activity in tumour cells. A still further object of the invention is to provide a method for treating tumour cells in a host in such a way that they become more sensitive to the above-mentioned alkylating agents.

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The present invention provides 6-hetarylalkyloxy pyrimidine derivatives of formula II:

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wherein

R is (i) a cyclic group having at least one 5- or 6- membered heterocyclic ring, optionally with a carbocyclic or heterocyclic ring fused thereto, the or each heterocyclic ring

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having at least one hetero atom chosen from 0, N or S, or a substituted derivative thereof; or

(ii) phenyl or a substituted derivative thereof, R^2 is selected from H, C_1 - C_5 alkyl, halogen or NH_2 , R^4 and R^5 which are the same or different are selected from H, NH-Y' or NO_n wherein Y' is H, ribosyl, deoxyribosyl, arabinosyl, R''XCHR''' wherein X is 0 or S and R'' is alkyl and R''' is H or alkyl, or substituted derivatives thereof, n=1 or 2, or R^4 and R^5 together with the pyrimidine ring form a 5-or 6-membered ring structure containing one or more hetero atoms,

with the proviso that R^2 is not NH^2 if R^4 and R^5 form a ring structure $1\dot{\text{X}}$



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wherein Y is H, ribosyl, deoxyribosyl, or R''XCHR''' wherein X is O or S, R'' and R''' are alkyl, or substituted derivatives thereof,

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and with the proviso that R is not phenyl in the following circumstances a) to h):

a) if R^2 and R^5 are NH_2 and R^4 is NO or NO_2

and pharmaceutically acceptable salts thereof,

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b) if R^2 is NH_2 and R^4 and R^5 form a ring structure X



X

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c) if R^2 is NH₂ and R^4 and R^5 form a ring structure XI

d) if ${\rm R}^2$ is ${\rm NH}_2$, and ${\rm R}^4$ is ${\rm NO}_2$ and ${\rm R}^5$ is H or ${\rm CH}_3$

10 e) if R^2 , R^4 and R^5 are NH_2 ,

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- f) if R^2 and R^5 are NH_2 and R^4 is H,
- g) if R^2 is H, and R^4 is NO_2 and R^5 is NH_2 , or
- h) if R^2 is F or OH, and R^4 and R^5 form a ring structure XII

Certain Q^6 -substituted guanine derivatives within the scope of the general formula in WO 94/29312 but not published therein have been found to have a surprisingly advantageous combination of properties which justifies the selection of such derivatives from among the class defined in WO 94/29312.

30 In another aspect, the present invention provides guanine derivatives of formula XIII:

wherein

E is 0 or S,

Y' is as defined for formula II above,

R⁶ is a cyclic group having at least one 5- or 6-membered heterocyclic ring, optionally with a carbocyclic or heterocyclic ring fused thereto, the or each heterocyclic ring having at least one hetero atom chosen from 0, N or S, or a substituted derivative thereof,

and pharmaceutically acceptable salts thereof, with the proviso that compounds published in WO 94/29312 are disclaimed.

In particular, the present invention selects advantageous compounds of formula XIV:

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wherein

 $\ensuremath{\mathsf{R}}^{10}$ is bromo, chloro or cyano, and

Y' is as defined for formula II.

25 Most preferably, R^{10} is bromo. A particularly preferred and selected compound is $\underline{0}^6$ -(4-bromothenyl)guanine having the formula XV:

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This compound has an advantageous combination of properties including potential for oral administration.

R or R^6 may suitably be a 5- or 6-membered heterocyclic ring or a benzo derivative thereof, in which latter case the pyrimidine moiety may be attached to R or R⁶ at either the heterocyclic or the benzene ring.

In preferred embodiments, R or R^6 is a 5-membered ring containing S or O, with or without a second ring fused thereto.

Preferably, R or R^{6} is a heterocyclic ring having at least 10 one S atom; more preferably, R or R⁶ is a 5-membered heterocyclic ring having at least one S atom; and most preferably, R or R^6 is a thiophene ring or a substituted derivative thereof. Alternatively, R or R⁶ may be a heterocyclic ring having at least one 0 atom, particularly, a 5-membered heterocyclic ring having at least one 0 15 atom and more particularly R or R^6 may be a furan ring or a substituted derivative thereof. As another alternative, R or R^b may be a heterocyclic ring having at least one N atom, particularly R or R^6 may be a 6-membered heterocyclic ring having at least one N atom and in particular, R or R^6 may be a pyridine ring. 20

The carbocyclic or heterocyclic ring fused to the heterocyclic ring in R or R^6 may itself be bicyclic e.g. naphthalene.

In general the term "substituted derivative" as used in 25 relation to any of the compounds of the invention means any substituted derivative whose presence in the compound is consistent with the compound having ATase depleting activity.

In the definition of Y or Y', the term "substituted derivative" includes further substitution by one or more of the following groups: hydroxy, halo, alkoxy, amino, alkylamino, amido or ureido. In a particularly preferred group of compounds, R" is hydroxy-substituted alkyl and R''' is H, so that Y' is hydroxyalkoxymethyl, preferably having 1 to 10 carbon atoms in the 35 alkoxy group.

- 11 -

In the definition of R or R^6 , the term "substituted derivative" includes substitution of the heterocyclic ring(s) and/or carbocyclic ring(s) by one or more of the following groups: alkyl, alkenyl, alkynyl, alkoxy, aryl, halo, haloalkyl, nitro, cyano, azido, hydroxyalkyl, SO_nR^7 where R^7 is alkyl and n=0,1 or 2, or a carboxyl or ester group of the formula $-COOR^8$ wherein R^8 is H or alkyl. Halo, haloalkyl, cyano, alkylenedioxy, SO_nR^7 (as defined above) and $-COOR^8$ wherein R^8 is alkyl are preferred substituents.

An alkyl, alkoxy, alkenyl, or alkynyl group preferably contains from 1 to 20, more preferably from 1 to 10 and most preferably from 1 to 5 carbon atoms. Halo includes iodo, bromo, chloro or fluoro. An aryl group preferably contains from 1 to 20, more preferably from 1 to 10 carbon atoms, particularly 5 or 6 carbon atoms.

One embodiment of the invention provides a pharmaceutical composition containing compounds of formula II or formula XIII, as defined above, or a pharmaceutically acceptable salt thereof, and a pharmaceutically acceptable excipient. Optionally the composition may also contain an alkylating agent such as a chloroethylating or methylating agent.

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In a further embodiment, the present invention provides a method for depleting ATase activity in a host comprising administering to the host an effective amount of a composition containing a compound of formula II or formula XIII as defined above, or a pharmaceutically acceptable salt thereof, more particularly a pharmaceutical composition as defined above. This method may alternatively be defined as a method of depleting ATase mediated DNA repair activity in a host.

The invention further provides a method for treating tumour cells in a host comprising administering to the host an effective amount of a composition containing a compound of formula II or

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formula XIII as defined above or a pharmaceutically acceptable salt thereof, more particularly a pharmaceutical composition as defined above and administering to the host an effective amount of a composition containing an alkylating agent. The method may be used for treatment of neoplasms including those which are known to be sensitive to the action of alkylating agents e.g. melanoma and glioma and others whose resistance to treatment with alkylating agents alone may be overcome by the use of an inactivator according to the invention.

The term "pharmaceutically acceptable salts" as used in this description and the claims means salts of the kind known in the pharmaceutical industry including salts with inorganic acids such as sulfuric, hydrobromic, nitric, phosphoric or hydrochloric acid and salts with organic acids such as acetic, citric, maleic, fumaric, benzoic, succinic, tartaric, propionic, hexanoic, heptanoic, cyclopentanepropionic, glycolic, pyruvic, lactic, malonic, malic, o-(4-hydroxy-benzoyl)benzoic, cinnamic, mandelic, methanesulfonic, ethanesulfonic, 1,2-ethanedisulfonic, 2-hydroxyethanesulfonic, benzenesulfonic, p-chlorobenzenesulfonic 2-naphthalenesulfonic, p-toluenesulfonic, camphorsulfonic, 4-methyl-bicyclo[2.2.2]oct-2-ene-1-carboxylic, glucoheptonic, 4,4'-methylenebis(3-hydroxy-2-naphthoic), 3-phenylpropionic, trimethyl-acetic, tertiary butylacetic, lauryl sulfuric, gluconic, glutamic, hydroxynaphthoic, salicylic, stearic, or muconic, and the like.

Subject to the provisos above the preferred compounds of the invention are those of:

Type 1
Formula III

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wherein:

R is as defined for formula II, particularly furyl or thienyl unsubstituted or substituted, preferably with a halogen such as chlorine, bromine or fluorine, or with cyano Y' is as defined for formula XIII, preferably Y' is H or HOCH₂CH₂OCH₂-;

 R^2 is H, NH_2 , C_1 - C_5 alkyl, preferably methyl, or halogen, preferably fluorine;

 R^3 is H or OH:

Type 2

Formula IV

wherein:

R is as defined for formula II, particularly phenyl, thienyl or furyl unsubstituted or substituted preferably with a halogen such as chlorine, bromine or fluorine, or with cyano, or phenyl having a methylenedioxy ring structure fused thereto; Y' is as defined for formula XIII;

25 X is CH or N; A is CH or N; and preferably when X = N, A = CH

Formula V

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wherein:

R is as defined for formula II

X is CH or N

A is CH or N;

Type 3
Formula VI

wherein:

R is as defined for formula II, particularly, thienyl or furyl unsubstituted or substituted preferably with a halogen such as chlorine or bromine;

Z is 0 or S or CH = CH;

A particularly preferred group of compounds of this type are $\underline{0}^6$ -(4-halothenyl)-8-thiaguanines, particularly $\underline{0}^6$ -(4-bromothenyl)-8-thiaguanine.

Formula VII

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wherein:

25 R is as defined for formula II;

U is CH or N;

V is CH or N;

W is CH or N;

provided that U, V and W are not all CH.

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Type 4

Formula VIII

wherein:

R is as defined for formula II, particularly thenyl or furyl optionally substituted with halogen preferably one or more of chlorine, bromine or fluorine;

T is H,
$$NH_2$$
 or NO_n where $n = 1$ or 2;
Q is H, NH_2 or NO_n where $n = 1$ or 2;

10 Type 5

Formula XVI

$$\begin{array}{c|c} RCH_2S \\ \hline N \\ NH_2 \\ \hline N \\ N \\ Y \\ \end{array} XVI$$

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wherein

20 R is as defined for formula XIII
Y' is as defined for formula II

Brief Description of Drawings

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The invention will be described in greater detail with reference to the accompanying drawings, in which:

Figures 1 to 4 are graphs showing the effect of pretreatment with compound B.4316 on Raji cell sensitization to different chemotherapeutic agents. Each graph plots percentage growth against the concentration ($\mu g/ml$) of the chemotherapeutic agent in the presence and absence of B.4316.

Figure 1 shows the effect of 1uM B.4316 pretreatment on Raji cell sensitization to temozolomide.

Figure 2 shows the effect of 10µm B.4316 pretreatment on Raji cell sensitization to BCNU.

5 Figure 3 shows the effect of 10µM B.4316 pretreatment on Raji cell sensitization to fotemustine.

Figure 4 shows the effect of $10\mu\text{M}$ B4316 pretreatment on Raji cell sensitization to melphalan and cisplatin.

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Figure 5 is a histogram showing the effect of 10 μ M B4316 pretreatment on Raji cell sensitization to different chemotherapeutic agents, measured as sensitization factor (SF, defined below) based on D $_{50}$ except for fotemustine where SF is based on D $_{80}$.

Figure 6 is a similar histogram showing the effect of $10\mu\text{M}$ B4349 pretreatment on Raji cell sensitization to different chemotherapeutic agents, with SF as for Figure 5.

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Figure 7 is a series of histograms showing the inactivation of ATase in A375M tumours and murine host tissues two hours after interperitoneal (i.p.) administration of various inactivator compounds at 5mg/kg. Inactivation was calculated as % of control ATase activity, measured as fm/mg protein.

Figure 8 is a graph showing the kinetics of ATase depletion and recovery in A375M tumours and murine host tissues after administration of B.4363 (20 mg/kg i.p.). The graph plots % of control ATase activity against time (hours).

Figure 9 is a graph of percentage residual activity of pure recombinant human ATase following incubation with increasing concentrations of inactivators $\underline{0}^6$ -benzylguanine (BeG), $\underline{0}^6$ -thenylguanine (B.4205) and $\underline{0}^6$ -(4-bromothenyl)guanine (B.4280). The line at 50% residual activity is used for calulating I_{50} values i.e. the concentration of inactivator required to

- 17 -

produce a 50% reduction in ATase activity. The I_{50} values shown are extrapolated from the curves. Preincubation was for 1 hour after which $[^3\mathrm{H}]$ -methylated substrate was added to determine residual activity of ATase.

Figure 10A is three graphs of percentage cell growth against temozolomide concentration (μ g/ml) showing the effect of pretreatment with BeG, B.4205 and B.4280 (0.5 μ M final concentration) on the sensitivity of Raji cells to the growth inhibitory effects of temozolomide. Inactivator or vehicle was given 2 hours prior to temozolomide.

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Figure 10B is a histogram for the inactivators of Figure 10A showing the sensitization factor based on D_{50} of Raji cells to growth inhibition by temozolomide.

Figure 11 is a histogram of ATase activity (fm/mg) against time (hours) showing the effect of ATase inactivators BeG, B.4205 and B.4280 on ATase activity in human melanoma xenografts grown in nude mice. Animals were given a single dose of the inactivators intraperitoneally (i.p.) at 30mg/kg or 60mg/kg and sacrificed after the times shown.

Figure 12 is a histogram showing the effect of ATase inactivators on ATase activity (fm/mg) in human melanoma xenografts grown in nude mice. Animals were given B.4205 or temozolomide alone or B.4205 or B.4280 in combination with temozolomide (50mg/kg)i.p. at the doses shown on three consecutive days (except where indicated) and sacrificed 24 hours after the final dose. The vehicles were corn oil for the inactivators and PBS (20%DMSO) for temozolomide.

Figure 13 is a histogram showing the effect of ATase
35 inactivators on ATase activity in livers of nude mice. Animals were
given the B.4205 or temozolomide alone or B.4205 or B.4280 in
combination with temozolomide (50mg/kg, i.p.) at the doses shown on

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three consecutive days (except where indicated) and sacrificed 24 hours after the final dose.

Figure 14A is a graph of % tumour growth against time (days) showing the effect of B.4205 on the sensitivity of human melanoma xenografts to growth inhibition by temozolomide. Animals were untreated, given temozolomide alone (100mg/kg, i.p.) or B.4205 (5, 10 or 20mg/kg i.p.) followed 1 hour later by temozolomide (100mg/kg, i.p.) on five consecutive days. Tumour growth was monitored as described. The data from a number of separate studies are presented.

Figure 14B is a graph of number of surviving mice against time (days) showing survival of animals (tumour-bearing nude mice) used in the study shown in Figure 14A. Groups of animals in which the xenografts had reached the maximum size were terminated.

Figure 15A is a graph showing the effect of B.4280 on the sensitivity of human melanoma xenografts to growth inhibition by temozolomide. Animals were untreated, given temozolomide alone (100mg/kg. i.p.) or B.4280 alone (20mg/kg, i.p.) or B.4280 (1, 5, 10 or 20 mg/kg, i.p.) followed 1 hour later by temozolomide (100mg/kg, i.p.) on five consecutive days. Tumour growth was monitored as described. The data from a number of separate studies are presented.

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Figure 15B is a graph showing the survival of the animals (tumour-bearing nude mice) used in the study shown in Figure 15A. Groups of animals in which the xenografts had reached the maximum size were terminated.

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Figure 16A is a graph of % tumour growth against time (days) showing the comparison of the effect of B.4280 given i.p. and orally (p.o.) on the sensitivity of human melanoma xenografts to growth inhibition by temozolomide. Animals were untreated, given temozolomide alone (100mg/kg) or B.4280 alone (20mg/kg, i.p.) or B.4280 (20mg/kg, i.p.) or B.4280 (30mg/kg, p.o.) followed 1 hour later by temozolomide (100mg/kg, i.p.) on five consecutive days.

WO 97/20843

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Tumour growth was monitored as described. The data from a number of separate studies are presented.

Figure 16B is a graph showing the survival of the animals used in the study shown in Figure 16A. Groups of animals in which the xenografts had reached the maximum size were terminated.

Figure 17 is a graph showing the survival of animals in a comparative test of the effects of BeG, B.4205 and B.4280 in 10 combination with temozolomide (TZ) in non-tumour-bearing DBA₂ mice. Animals were given temozolomide alone (100mg/kg i.p) or BeG (10 or 20mg/kg i.p.), B.4205 (10 or 20 mg/kg i.p.) or B.4280 (10 or 20 mg/kg i.p.) followed one hour later by temozolomide (100mg/kg i.p.) on five consecutive days. 15

Figures 18 to 21 consist of pairs of graphs showing the kinetics of ATase depletion and recovery in various tumours and murine host tissues after administration of B.4280 at the doses indicated. The graphs plot ATase activity (fm/mg protein) and % of control ATase activity against time (hours):

Figure 18 relates to B.4280 (20 mg/kg i.p.) in A375M tumours and other tissues.

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Figure 19 relates to B.4280 (30mg/kg p.o.) in A375M tumours and other tissues

Figure 20 relates to B.4280 (30mg/kg i.p.) in MCF-7 tumours and other tissues. 30

Figure 21 relates to B.4280 (20mg/kg i.p.) in DU-145 tumours and other tissues.

Figure 22 is a graph of % tumour growth against time (days) 35 showing the effect of B.4280 on the sensitivity of MCF-7 tumours to growth inhibition by temozolomide. Animals were untreated, were

given temozolomide alone (100 mg/kg, i.p.) or B.4280 (PaTrin-2) (20mg/kg i.p.) alone, or B.4280 (20mg/kg i.p.) followed 1 hour later by temozolomide (100mg/kg i.p.) on five consecutive days.

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Figure 23 consists of graphs of % tumour growth, number of surviving mice and mean weight (g) against time (days) showing the effect of a single dose of B.4280 (PaTrin-2) on the sensitivity of melanoma tumours to growth inhibition by a single dose of fotemustine. Animals were given fotemustine (20mg/kg i.p.) alone, or B.4280 (30mg/kg p.o) followed 1 hour later by fotemustine (20mg/kg i.p.).

Figure 24 consists of graphs of % tumour growth and number of surviving mice against time (days) for sensitization of A375M tumours with B.4205 (PaTrin-1) and B.4280 20mg/kg pretreatment followed by 150mg/kg temozolomide using a 5 day schedule as for Figure 22.

Figure 25 consists of graphs of % tumour growth, number of surviving mice and mean weight (g) against time showing sensitization of A375M tumours to temozolomide (100mg/kg i.p.) following administration of 20mg/kg B.4349 or B.4351 (i.p.).

25 Figure 26 is a figure showing ATase activities in A375M tumours and murine host tissues at 2 hours and 24 hours following i.p. administration of 90mg/kg B.4335.

In the specification the abbreviations "1h" or "2h" etc. mean 30 "1 hour", "2 hours" etc.. In the drawings the abbreviations "Temo" and "Tz" refer to temozolomide.

Figure 27 consists of graphs of % tumour growth and weight (% of day 1 value) against time (days) showing tumour DU-145 prostate

35 xenograft growth after temozolomide (100mg/kg/day) and/or B.4280
(PaTrin-2)(20mg/kg/day) days 1-5. Points are the means of values from at least 4 mice. Growth delays in each group were (p value):

PaTrin-2 alone 0.1 days (>.05); temozolomide alone 7.8 (>.05). Both agents 15.3 (0238).

Figure 28 is a reaction scheme for synthesis of 5 $0^6[^3H]$ - (4-bromothenyl) guanine.

Figure 29 shows co-chromatography of authentic B.4280 and readjoactivity in the product of $0^6 - [^3H] - (4-bromothery)$ guanine synthesis. Shading indicates counts recovered (LH axis) and the line OD at 254nm (RH axis).

Figure 30 shows transfer of radioactivity from $0^6 - [^3H] - (4-bromothenyl)$ guanine to rhATase after one hour incubation at 37°C. 15

Description of the Preferred Embodiments

Examples of compounds of the invention are shown in Tables 1a 20 and 1b. They were synthesized by the procedures presented below, adapted as appropriate.

Type 1

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- 0^6 -Substituted hypoxanthines were made by the action of 25 Α. alkoxide RCH₂ONa on the quaternary salt $\underline{\mathtt{N}},\underline{\mathtt{N}},\underline{\mathtt{N}}$ -trimethyl-1 $\underline{\mathtt{H}}$ -purin-6-aminium chloride. 1
- 0^6 -Substituted 2-methylhypoxanthines were made similarly, В. from the quaternary salt from diazabicyclooctane (DABCO) and 30 6-chloro-2-methylpurine.²
- $\underline{0}^6$ -Substituted 2-fluorohypoxanthines were made by С. diazotisation of the corresponding guanines using sodium nitrite and concentrated fluoboric acid at -25°C.3 35

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- D. Q⁶-Substituted 9-(2-hydroxyethoxymethyl)guanines were made by condensing the corresponding guanines after silylation with 2-acetoxyethoxymethyl bromide in the presence of mercuric cyanide followed by saponification of the Q-acetyl group. 4
- E. <u>0</u>⁶-Substituted 8-hydroxyguanines were made from 6-hetarylmethyl-2,4,5-triaminopyrimidines and 1, 1-carbonyldimidazole in DMF. Reaction of 6-chloro-2,4-diaminopyrimidine with alkoxide in DMSO, followed by nitrosation with sodium nitrite in aqueous acetic acid and reduction using sodium hydrosulphite in aqueous DMF, gave the 2, 4, 5-triamines.

15 <u>Type 2</u>

- A. 0^6 -Substituted 8-azaguanines were made from the above triamines and sodium nitrite in aqueous acetic acid.
- 20 B. $\underline{0}^6$ -Substituted 8-aza-7-deazaguanines were made from the alkoxide RCH₂ONa and 2-amino-6-chloro-8-aza-7-deazapurine⁷ in sulfolane or from the DABCO quaternary salt (in DMSO solvent) derived from it.

25 <u>Type 3</u>

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- A. $\underline{0}^6$ -Substituted 8-oxaguanines were made by lead tetraacetate oxidation of 6-hetarylmethyl-2,4-diamino-5-nitrosopyrimidines obtained as under Type IE.
 - B. $\underline{0}^6$ -Substituted 8-thiaguanines were made from the triamine intermediates under Type IE and N-tosylthionylimine in pyridine.
- C. $\underline{0}^4$ -Substituted pterins were made from these triamines and glyoxal with sodium metabisulphite.

- 23 -

Type 4

A and B.

- 5 These pyrimidines were obtained as under Type IE.
 - C. <u>0</u>⁶-Substituted 2,4-diamino-5-nitropyrimidines were made by the action of alkoxide RCH₂ONa in DMSO on 6-chloro-2,4-diamino -5-nitropyrimidine. 11

Type 5

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 \underline{S}^6 -Substituted 6-thioguanines were prepared from the thiolate RCH₂SNa and the quaternary salt 2-amino-N,N,N-trimethyl-1 \underline{H} -purin-6-aminium chloride (WO 94/29312).

 0^6 -Substituted guanines as listed in Tables 6a and 6b were made by the standard preparation as described in WO 94/29312, usually with 3mmol alcohol RCH₂OH per mmol quaternary salt.

The alcohols were made as described in WO 94/29312 by sodium borohydride reduction of the corresponding aldehydes, with two exceptions. For 4-bromothenyl alcohol 12 required for B.4280 the aldehyde is commercially available. 5-Chlorothiophen-2-aldehyde 13 and 5-methylthiothiophen-2-aldehyde 14 were prepared by Vilsmeier reaction on 2-chlorothiophen and 2-methylthiothiophen respectively. Sodium borohydride reduction of the methylthioaldehyde followed by sodium periodate oxidation 15 of the resulting methylthioalcohol yielded the methylsulphinylalcohol required for B.4294. Reduction of the chloroaldehyde gave 5-chlorothenyl alcohol 16 for B.4281.

Several other aldehydes were obtained by halogenation of the appropriate thiophen aldehyde or furfural. Thus, direct bromination gave 5-bromofurfural and thence the alcohol for B.4336. Halogen in presence of aluminium chloride on thiophen-2-aldehyde yielded 4-chlorothiophen-2-aldehyde (for the alcohol for

B.4298), on thiophen-3-aldehyde yielded 2-bromothiophen-4-aldehyde²⁰ (and eventually B.4313), and on 5-chlorothiophen-2-aldehyde yielded 4,5-dichlorothiophen-2-aldehyde²¹ (for the alcohol²² for B.4318).

Cyanoaldehydes were obtained from copper cyanide and the corresponding bromoaldehydes in refluxing dimethylformamide. 5-Cyanothiophen-2-aldehyde and its 4-cyano isomer then gave the 5-cyano and 4-cyano alcohols, for B.4283 and B.4317 respectively.

4-Methoxythenyl alcohol²⁶ (for B.4300) was prepared as described from 2,3-dibromosuccinic acid and methyl thioglycollate, and ultimate reduction of the methyl ester (not aldehyde in this case) by lithium aluminium hydride and 2-chloro-4-picolyl alcohol²⁷ (for B.4321) by sodium borohydride reduction²⁸ of the corresponding acid chloride, made in turn from reaction²⁹ of phosphorus oxychloride/pentachloride on isonicotinic acid N-oxide.

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For B.4282, 3-pyridinemethanol N-oxide is commercially available. 5-Methylsulphonylthenyl alcohol (for B.4309) was obtained by m-chloroperbenzoic acid (MCPBA) oxidation of the alcohol resulting from reduction of 5-methylthio-2-thiophenecarboxaldehyde 30.

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6-Chloro-3-pyridinemethanol (for B.4319) and 5-bromo-3-pyridinemethanol (for B.4320) were made by treatment of 6-chloro and 5-bromonicotinic acids respectively with phosphorus oxychloride/pentachloride and reduction of the resulting acid chlorides with sodium borohydride²⁸. Isothiazole-4-methanol (for B.4354) was obtained by reduction of the corresponding methyl ester (A. Adams and R. Slack, <u>J. Chem. Soc.</u> 1959, 3061) with lithium aluminium hydride (M. Hatanaka and T. Ishimaru, <u>J. Med. Chem.</u> 16, 1973, 978).

4-bromo-2-thiophenecarboxaldehyde was converted into the

4-lithio derivative (A.L. Johnson, J. Org. Chem. 41, 1976, 1320) of its ethylene acetal and reaction of this organometallic with dimethyl disulphide followed by acid hydrolysis gave 4-methylthio-2-thiophenecarboxaldehyde (R. Noto, L. Lamartina, C. Arnone and D. Spinelli, J. Chem. Soc., Perkin Trans, 2, 1987, 689). Sodium borohydride reduced this aldehyde to the 4-methylthio alcohol (for B.4356), which in turn with one of two equivalents of MCPBA yielded the 4-methylsulphinyl and 4-methylsulphonyl alcohols (for 10 B.4377 and B.4361 respectively). Reaction of the above organometallic with naphthalene-2-sulphonyl azide (A.B. Khare and C.E. McKenna, Synthesis, 1991, 405) and sodium pyrophosphate followed by hydrolysis by the method (P. Spagnolo and P. Zanirato, <u>J. Org. Chem.</u>, 43, 1978, 3539) for the preparation of other azidothiophene aldehydes gave 4-azido-2-thiophenecarboxaldehyde 15 leading to the alcohol for B.4373.

5-Iodo-3-thiophenemethanol (for B.4357) came from the aldehyde obtained by treatment of 3-thiophenecarboxaldehyde with iodine-iodic acid-sulphuric acid (R. Guilard, P. Fournari and M. Person, <u>Bull. Soc. Chim. France</u>, 1967, 4121).

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2-Naphtho[2, 1-b]thienylmethanol (for B.4366) was prepared by lithium aluminium hydride reduction of the corresponding carboxylic acid (M.L. Tedjamulia, Y. Tominaga, R.N. Castle and M.L. Lee, J. Heterocycl. Chem., 20, 1983, 1143). 5-Phenylthenyl alcohol (m.p. 91.5°C, for B.4378) resulted from sodium borohydride reduction of the aldehyde (P. Demerseman, N.P. Buu-Hoi and R. Royer, J. Chem. Soc., 1954, 4193) obtained by Vilsmeier reaction of 2-phenylthiophene (from Gomberg-Bachmann reaction (N.P. Buu-Hoi and N. Hoan, Rec. trav. chim., 69, 1950, 1455) of benzenediazonium chloride and alkali with thiophene).

By way of specific example, the preparation of $\underline{0}^6$ -(4-bromothenyl)guanine (B.4280) will now be described.

Preparation of 0^6 -(4-bromotheny1)guanine A solution of 4-bromothenyl alcohol 12 [4.63g, 24mmol; R_f 0.38 in TLC(PhMe-MeOH, 4:1)] in DMSO (4ml) was treated cautiously with sodium hydride (60% in oil; 0.64g, 16mmol). After 1 hour's stirring, 2-amino-N,N,N-trimethyl-1H-purin-6-aminium chloride (1.83g, 8mmol) was added. After 1 hour's further stirring, acetic acid (1.3ml) followed by ether (240ml) was added and the solid filtered off after 1-2h. Removal of solvents and excess of alcohol (b.p. $85-90^{\circ}$ C/0.4mm) from the filtrate yielded a negligible second fraction (17mg). The main crop was triturated with water (10ml), affording substantially pure product (1.89g, 73%) with $\rm R_f$ 0.22 in TLC (PhMe-MeOH, 4:1). It was recrystallized by dissolving in hot methanol (100ml) and then concentrating. Analytical data are given in Tables 6a and 6b, together with data for other compounds. Other 15 typical synthetic procedures are described by way of example in a special section later in this text.

Compounds of formula II or XIII in which Y' is R''XCHR''' and R''' is alkyl (seco-nucleosides) may be prepared by an analogous preparation to the reaction of Obenzylguanine with -chloro-ethers (MacCoss et al., Tetrahedron Lett.; European Patent Application No. 184,473., loc. cit.) or with alkyl bromides (e.g. Kjellberg, Liljenberg and Johansson, Tetrahedron Lett., 1986, 27, 877; Moschel, McDougall, Dolan, Stine, and Pegg, J.Med. Chem., 1992, 35, 4486).

Typical "sugar" components corresponding to R''XCHR''', leading to seco-nucleosides, are made by methods described in <u>e.g.</u> McCormick and McElhinney, <u>J. Chem. Soc. Perkin Trans. 1</u>, 1985, 93; Lucey, McCormick and McElhinney, <u>J. Chem. Soc. Perkin Trans. 1</u>, 1990, 795.

Compounds of formula II or XIII in which Y is ribosyl or deoxyribosyl (nucleosides) may be prepared by methods analogous to the syntheses of <u>0</u>⁶-benzylguanine riboside and 2-deoxyriboside (Moschel <u>et al</u>. 1992; cf. Gao, Fathi, Gaffney <u>et al</u>., <u>J. Org. Chem.</u>,

- 27 -

1992, **57**, 6954; Moschel, Hudgins and Dipple, <u>J. Amer. Chem. Soc.</u>, 1981, **103**, 5489) (see preparation of Ribosides above).

5 Industrial Applicability

The amount of the compound of the present invention to be used varies according to the effective amount required for treating tumour cells. A suitable dosage is that which will result in a concentration of the compound of the invention in the tumor cells to be treated which results in the depletion of ATase activity, e.g. about 1 - 2000 mg/kg body weight, and preferably 1 - 800 mg/kg body weight, particularly 1-120 mg/kg body weight, prior to chemotherapy with an appropriate alkylating agent.

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The pharmaceutical composition of the invention may be formulated in conventional forms with conventional excipients, as described for example in WO 91/13898 and WO 96/04281 and U.S. Patents 5,091,430 and 5,352,669, the contents of which are incorporated herein by reference in their entirety. The composition may contain the inactivator according to the invention together with an appropriate alkylating agent; or the composition may comprise two parts, one containing the inactivator and the other containing the alkylating agent. The method of administering the compounds of theinvention to a host may also be a conventional method, as described in WO 91/13898 for example. For administration of an inactivator according to the invention to patients, the pharmaceutical composition may suitably contain the inactivator in a suitable vehicle such as 40% polyethyleneglycol 400 in saline solution, or in saline or 3% ethanol (in saline), for intravenous injection, or in a powder form in suitable capsules for oral administration.

Alkylating agents may be administered in accordance with known techniques and in conventional forms of administration, as described in WO 91/13898 for example or preferably as a single dose immediately after or up to 24 hours after but preferably around 2

hours after administration of the ATase inactivating agents and also at doses lower than those used in standard treatment regimen. A reduction in dose may be necessary because the inactivators would generally be anticipated to increase the toxicity of the alkylating agents. Examples of chloroethylating agents include 1,3 bis (2-chloroethyl)-1-nitrosourea (BCNU). 1-(2-chloroethyl)-3-cyclohexyl-1-nitrosourea (CCNU), fotemustine, mitozolomide and clomesone and those described in McCormick, McElhinney, McMurry and Maxwell J. Chem. Soc. Perkin Trans. I, 1991, 10 877 and Bibby, Double, McCormick, McElhinney, Radacic, Pratesi and Dumont Anti-Cancer Drug Design, 1993, 8, 115. Examples of methylating agents include temozolomide (British Patent GB 2 104. 522 and U.S. Patent 5,260,291 the contents of which are incorporated herein in their entirety) and dacarbazine, procarbazine, and 15 streptozocin.

<u>Methods</u>

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$20 ext{0}^6$ -alkylguanine-DNA-alkyltransferase assay

Varying amounts of recombinant ATase or cell/tissue extracts were incubated with $[^3H]$ -methylnitrosourea-methylated calf thymus DNA (specific activity, 17Ci/mmol) at 37°C for 1 hour in a total volume of 30Qul buffer I/[50mM Tris/HCl (pH8.3), 3mM dithiothreitol (DTT), 1mM EDTA] containing 1mg/ml bovine serum albumin (IBSA) for recombinant ATases and tissue extracts, or 1.1ml buffer I for cell extracts. After incubation, bovine serum albumin (100ml of 10mg/ml in buffer I) and perchloric acid (100µl of 4M perchloric acid for 300µl volumes and 400µl for 1.1ml volumes) and 2ml of 1M perchloric acid were added. Samples were then heated at 75°C for 50 minutes to hydrolyze the DNA. Samples were then centrifuged at 3,000rpm for 10 minutes and the precipitate washed once with 4ml of 1M perchloric acid, before being resuspended in 300µl of 0.01M sodium hydroxide and dissolved in 3ml of aqueous scintillation fluid (Ecoscint A, National Diagnostics). Counting efficiency was approximately 30%. ATase specific activity was calculated from the region where the

- 29 -

activity was proportional to the amount of extract added, since with higher amounts of extracts the reaction becomes substrate limiting. ATase activity is expressed as fmol methyl transferred to protein per mg of total protein in the extract.

Method of Purification of Recombinant ATases

The cDNA cloning and overexpression of the human ATase has been reported previously 30. Purification of the recombinant 10 proteins was achieved either by affinity chromatography through a DNA-cellulose column as described by Wilkinson et al., 31, 32, or by DEAE-cellulose ion-exchange chromatography. For the latter, the ATase protein was partially purified by ammonium sulphate 15 precipitation (30 - 60%) and dialyzed against 10 mM Tris-HCl pH 7.5. 1 mM DTT, 2 mM EDTA, 10% glycerol, before loading on a DEAE-cellulose column. The ATase was then eluted with a 0-0:1 M NaCl gradient. The purified human ATase protein retained activity for more than one year when stored at high concentration at -20°C in buffer I [50 mM-Tris/HCl (pH 8.3)/3 mM-dithiothreitol/1 mM-EDTA] 20 and could be thawed and refrozen several times without substantial loss of activity.

Incubation with Inactivators and ATase assay

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Compounds to be tested were dissolved in DMSO to a final concentration of 10 mM and diluted just before use in buffer I . Recombinant ATase was diluted in buffer I containing 1 mg/ml bovine serum albumin (IBSA) and titrated as described above in order that the reaction be conducted under ATase, and not substrate, limiting conditions. In each assay, fixed amounts of ATase (60-75 fmol) were incubated with varying amounts of 0^6 -benzylguanine, or test compound in a total volume of 200 µl of IBSA containing 10 µg of calf thymus DNA at 37° C for 1 hour. The $[^3$ H]-methylated-DNA substrate (100 µl containing 4 µg of DNA and 100 fmol of 0^6 -methylguanine) was then added and incubation continued at 37° for 1 hour, until the reaction was complete. Following acid

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hydrolysis of the DNA as described above the $[^3H]$ -methylated protein was recovered and quantitated by liquid scintillation counting. I_{50} is the concentration of inactivator required to produce a 50% reduction in ATase activity under the above conditions.

Cell Culture and preparation of extracts

Mammalian cells including Raji cells (a human lymphoblastoid cell line from a Burkitt's lymphoma), A375M cells (human melanoma cells), MCF-7 cells (human breast cancer cells) and PC3 and DU145 (both human prostate cancer cells) were cultured under standard conditions. For example, Raji cells were grown in suspension culture in RPMI medium supplemented with 10% horse serum. Cell pellets were resuspended in cold (4°C) buffer I containing 2 ug/ml leupeptin and sonicated for 10 seconds at 12 um peak to peak distance. After cooling in ice, the cells were sonicated for a further 10 seconds at 18 um. Immediately after sonication, 10 ul/ml of phenylmethanesulphonylfluoride (PMSF 8.7 mg/ml in 100% ethanol) was added and the sonicates centrifuged at 15 000cpm for 10 minutes at 4°C to pellet cell debris. The supernatant was transferred to a tube on ice and kept for determination of ATase activity (see above).

Stability of Inactivators at 37°C by Spectrophotometry.

Inactivators (10mM in DMSO) were diluted to 0.1mM in prewarmed degassed PBS (pH 7-7.2). PBS (Phosphate buffered saline) is 0.8% NaCl, 0.02% KCl, 0.15% Na $_2$ H $_2$ PO $_4$, 0.02% KH $_2$ PO $_4$. Samples were immediately transferred to a CARY13 spectrophotometer (cuvette block held at 37 $^{\circ}$ C) and scanned at an appropriate wavelength (according to the spectral properties of the compound) at 5-10 minute intervals for up to 80 hours. The results were expressed as percentage absorbance change versus time and T1/2 values (half life) extrapolated from this. In the tables the results of these tests are identified by "in PBS" or "by Spec".

Stability of Inactivators by ATase Assay

Inactivators (10 μ m in DMSO) were diluted to the appropriate concentration (I $_{Q\Omega}$ calculated from previous I $_{S\Omega}$ determination

- 31 -

data) in buffer I without DTT and incubated for varying times at 37° C. Samples were then taken for use in the competition assay to assess the compound's ability to inactivate human ATase. The results were expressed as reduction in inactivating activity versus time and T 1/2 values extrapolated from this.

Inactivation of ATase activity in Raji cells.

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Raji cells were diluted to between $5 \times 10^5/\text{ml}$ and $10^6/\text{ml}$ in medium containing either the appropriate concentration of inactivator or an equivalent volume of vehicle (DMSO). Following incubation at 37°C for 2 hours the cells were harvested by centrifugation, washed twice with PBS and the resulting cell pellets (between 5×10^6 and 10^7 cells per pellet) stored at -20°C . ATase activity was determined as described above, in duplicate cell extracts and expressed as the percentage activity remaining, based on that present in the untreated controls (350-450 fm/mg depending on the assay). I_{50} (i.e concentration of inactivator required to reduce ATase activity by 50%) values were extrapolated from this data.

Sensitization of Mammalian cells to Cytotoxic Agents.

Sensitization of mammalian cells to the cytotoxic effects of BCNU, temozolomide and other cytotoxic agents following a 2 hour pretreatment with inactivator was analysed using an XTT-based growth inhibition assay 22 . Cells were plated in 96 well plates (for example in the case of Raji cells at 500 cells/well) and incubated at $37^{\rm O}{\rm C}$ for 30 minutes prior to the addition of medium containing either the appropriate concentration of inactivator or an equivalent volume of vehicle. Following a 2 hour incubation at $37^{\rm O}{\rm C}$, medium containing either increasing doses of cytotoxic agent or equivalent vehicle was added and the cells allowed to grow for 6 days. At this time XTT solution was added and the cells incubated for a further 4 hours at $37^{\rm O}{\rm C}$. The resulting red/orange formazan reaction product was quantified by measuring absorption at 450nm on a microtitre platereader.

From this data the percentage growth of cells relative to that in control wells was determined for a range of BCNU, temozolomide or other cytotoxic agent doses in both the presence and absence of inactivator. Sensitization factor (SF) based on D_{50} (D_{50} . $^{\rm C}/D_{50}$. $^{\rm I}$) was determined by dividing the D_{50} (i.e. dose at which there was 50% growth versus the controls untreated with alkylating agent) calculated for the cytotoxic agent alone (D_{50} . $^{\rm C}$) by that for the cytotoxic agent plus inactivator (D_{50} . $^{\rm I}$). A value of one (1) thus indicates no sensitization by the inactivator. Comparable Sensitization factors were also determined in some cases based on D_{60} and D_{80} , i.e. the dose at which there was respectively 60% or 80% growth compared to the untreated controls. In Table 3 the Sensitization Factor D_{50} . $^{\rm C}/D_{50}$. I is shown as D_{50} control / D_{50} 'B', with the letter 'B' referring to the inactivator compound.

Xenograft Studies

20 Animals

Swiss mouse derived athymic male mice (o/nu) weighing between 20-30g were obtained from ZENECA Pharmaceuticals, Alderley Park, Macclesfield, Cheshire, SK10 4T6, England. Animals were housed 4-5/cage in filter top cages and had access to food and water ad libitum. All animals were maintained under a controlled 12h-light-12h-dark cycle. These animals were used for all tests except those which are shown in Figures 11 and 17 and Table 8, as mentioned below.

30 Cells

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A375M (human melanoma) and DU145 (human prostate cancer) cells were grown in DMEM containing 10% foetal bovine serum (FBS). MCF-7 (human breast cancer cells) were grown in DMEM containing 10% FBS supplemented with 100iu insulin.

Tumours

A375M, DU145 and MCF-7 cells (10^6) in 100 μ l PBS were

injected subcutaneously into the right-hand flank of 8-10 week old o/nu athymic mice. These cells were allowed to develop into a tumour for 3-4 weeks (A375M and DU145 cells) and 4-6 weeks (MCF-7 cells). Once established, tumours were maintained by subcutaneous implantation of 2mm³ blocks into the right-hand flank of athymic o/nu mice. MCF-7 tumours are oestrogen postive and require oestrogen for growth. This was supplied as a subcutaneous implant (see below) at the tail base simultaneously to the tumour implant and monthly thereafter.

Preparation of Oestrogen Pellets

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468mg **\mathbb{\beta}**-oestradiol was added to 9.7g silastic and mixed until evenly distributed. '1.1g of curing agent was added and the whole mixture spread into 3 (26mm x 12mm x 1mm) glass fomers. These were then incubated at 42°C overnight before being cut into 2mm x 2mm x 1mm cubes, so that each pellet contained 2mg estradiol.

ATase Depletion Experiments

Tumours were implanted as previously described and left 3-6 weeks to establish depending on tumour type. An inactivator was homogenized in corn oil at 5mg/ml before administration by interperitoneal injection (i.p.) or oral gavage (p.o.). Mice were sacrificed at various times up to 72h and tumours and murine tissues taken for ATase assay. Samples were snap frozen and stored at -20°C until analysis.

Tumour Sensitization Experiments

O/nu mice were treated with the appropriate dose of the inactivator as indicated (4mg/ml in corn oil) or the appropriate vehicle as a control 1 hour prior to administration of the appropriate dose of the cytotoxic agent (e.g.temozolomide 6mg/ml in PBS + 20% DMSO) or fotemustine or BCNU (2mg/ml in PBS + 3% ethanol) using the doses and schedules indicated.

Tumour Measurements

Animals were weighed twice weekly and xenograft tumour

measurements taken using digital calipers. Tumour volume was calculated using the formula $(h \times w \times 1) \pi / 6$. Measurements continued until the tumour reached the maximum allowable volume (i.e. 1cm^3), Results were expressed as percentage tumour growth using day 1 tumour volumes as controls.

In the tests on the compounds shown in Table 6 and in Figures 9 to 17, the Methods used were as described in WO 94/29312. The following items a) to c) are also to be noted:

a) Standard ATase assay

ATase substrate DNA was prepared by incubation of purified calf thymus DNA with N-[3 H]-methyl-N-nitrosourea (18.7 Ci/mmole, Amersham International). Cell or tissue extracts were incubated with [3 H]-methylated-DNA substrate (100yl containing 6.7µg of DNA and 100fmol of 6 -[3 H]methylguanine) at 37°C for 60 mins.. Following acid hydrolysis of the DNA as previously described 33 the [3 H]-methylated protein was recovered and quantitated by liquid scintillation counting.

b) Drug Treatment

Mice were treated with the inactivator as a suspension in corn oil by intraperitoneal injection (i.p.) or by oral gavage (p.o.) 60 mins prior to temozolomide (100mg/kg in 20%DMSO in phosphate-buffered saline) which was always given by intraperitoneal injection: this schedule was repeated on days 1 to 5 inclusive. Controls received vehicle alone, inactivator alone or temozolomide alone.

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c) Animals

The mice in the tests shown in Figure 11 and Table 8 were BALB-C derived athymic male mice (nu/nu athymic) from the in-house breeding colony of the Paterson Institute for Cancer Research as described in WO 94/29312 (Animal Services Unit-ASU Mice).

The mice in the tests shown in Figures 12-16 were Swiss mouse derived athymic male mice (o/nu athymic) as described above.

The mice in the tests shown in Figure 17 were DBA₂ mice from the in-house breeding colony of the Paterson Institute for Cancer Research (Animal Services Unit), originally from the Jackson Laboratory in 1970.

10 Test Results

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The results of the ATase depletion assay on the compounds of Table 1 are shown in Table 2 or Table 3. Many of the compounds tested were more efficient in inactivating ATase than $\underline{0}^6$ -benzylguanine. In accordance with the results in WO 94/29312 the parent application, compounds in which R is a heterocyclic group were more efficient than the comparable compounds having benzyloxy side chains. In general the compounds in which RCH₂ is substituted or unsubstituted thenyl were the most efficient, the most preferred being halo-substituted thenyl having its halo substituent in a 1,3-relationship with the methyleneoxy group attached to the pyrimidine residue.

Tables 3, 4 and 5 summarize data for a number of parameters.

Table 3 includes depletion assay results for recombinant ATase of the following types:

hAT = human mAT = mouse

rAT = rat

30 chAT = Chinese hamster

ogt = <u>E. Coli</u> ogt gene

ada = <u>E. Coli</u> ada gene

The combinations of properties for the various inactivators

35 can be seen in the tables. The following surprising points are
noted in particular:

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B.4316 is a compound of surprisingly high water solubility. B.4335 is a compound that is unexpectedly much more effective in the inactivation of ATase in Raji cells than of pure recombinant protein: generally, the I_{50} for inactivation of recombinant ATase *in vitro* is lower or similar to that in cultured cells.

B.4343 is a compound that has a very low I_{50} for ATase in vitro but is not as capable as agents with higher I_{50} s (e.g. B.4335) in the sensitization of Raji cells to the growth inhibitory effects of temozolomide. A similar example is B.4351 versus B.4349.

B.4316 was twice as effective as B.4280 but sensitization to temozolomide of Raji cells was almost identical. Thus different cell lines may respond surprisingly differently to these agents.

Figures 1 to 3 show that temozolomide, BCNU and fotemustine inhibit the growth of Raji cells in a dose-dependent manner but sensitivity is greatly increased by exposure to B.4316 at 0.1, 1.0 and 10µM respectively. In contrast B.4316 had no measurable effect on growth inhibition of Raji cells by melphalan or cisplatin (Fig. 4). This indicates that the inactivators were specifically sensitizing cells to the Q^6 -alkylating agents and not other classes of alkylating compound.

Figures 5 and 6 respectively show the B.4316 and B.4349 sensitization factors for the above therapeutic agents in Raji cells.

Figure 7 shows that of the inactivators examined human melanoma xenograft ATase depletion was complete only after administration of B.4314 and B.4351 under the experimental conditions used. The former was more effective in ATase depletion in liver and kidney of host animals whilst the latter was more effective in the brain, suggesting its relative ease in passing the blood-brain barrier. Noteworthy is the fact that whilst B.4311 was one of the most effective agents in sensitizing Raji cells to the

PCT/IE96/00084

toxic effects of temozolomide, it was surprisingly one of the least effective agents in depleting mouse tissue or tumour xenograft ATase activity.

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Figure 8 shows that B.4363 depletes ATase more effectively in human melanoma xenografts than in murine host tissues under the conditions used: relatively little effect was seen in brain tissue, suggesting its poor ability to cross the blood brain barrier.

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The test results for the compounds of Table 6 (and some in Table 1) are shown in Table 7 and Figures 9 to 27.

B.4280, which is $\underline{0}^6$ -(4-bromothenyl)guanine and has its bromo substituent in a 1, 3-relationship with the methylene group attached to the guanine residue, was more efficient in inactivating ATase in vitro than its 5-bromo analogue B.4269, in which the bromo substituent is in a 1, 4-relationship with the methylene group. Both B.4280 and B.4269 were more efficient than the unsubstituted thenyl derivative B.4205 despite having considerably larger $\underline{0}^6$ substituents.

Another preferred compound is B.4317 which is $\underline{0}^6$ -(4-cyanothenyl)guanine. B.4317 is a more efficient inactivator in vitro than its 5-cyano analogue B.4283 or the unsubstituted thenyl derivative B.4205.

Typical ATase inactivation profiles for BeG and B.4205 and B.4280 are shown in Fig. 9.

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The inactivation of ATase resulted in the sensitization of Raji cells to the growth inhibitory effects of temozolomide (Fig. 10). B.4280 was considerably more effective than either B.4205 or BeG in this respect.

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ATase in human melanoma xenografts was inactivated by BeG, B.4205 and B.4280 (Fig. 11) with some indication that the rates of

recovery of ATase activity were different between the agents. B.4280 was the most effective *in vivo* inactivator at the doses examined.

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B.4280 was able to inactivate ATase in most tissues as shown in Table 8. Thus, activity in brain, testis and bone marrow was near to control levels by 24 hours whereas lung and spleen activity had not completely recovered by 48 hours. Tumour activity was very low at 24 hours but had recovered completely by 48 hours. Differential recovery rates might be an important factor in the toxicity of ATase inactivators when used in combination with CNU or temozolomide.

15 Combinations of B.4205 or B.4280 and temozolomide given over three days were more effective in ATase inactivation in tumour xenografts than either agent alone (Fig. 12). Decreasing the dose of B.4205 had no major effect on the ability of the agent to inactivate ATase, 10mg/kg being as effective as 60mg/kg. B.4280 was more 20 effective than B.4205 at equivalent doses. As before (Fig. 11) there was some indication that ATase recovery was less efficient in the tumour xenograft (Fig. 12) than in the liver (Fig. 13).

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WO 97/20843 PCT/IE96/00084

B.4205 (Fig. 14A) and B.4280 (Fig. 15A) were effective in sensitizing human melanoma xenografts to the growth inhibitory effects of temozolomide. A comparison of the two sets of data indicates that B.4280 was about twice as effective as B.4205 in this respect. At equi-effective doses for tumour growth inhibition, B.4280 seems to be less toxic than B.4205 (Figs. 14B and 15B).

In experiments using DBA₂ mice in combination with BCNU,

B.4280 was considerably less acutely toxic than B.4205 or BeG as shown in Table 9. Oral administration of B.4280 was shown to be almost as effective as i.p. administration in sensitizing human melanoma xenografts to the growth inhibitory effects of temozolomide (Fig. 16A). Furthermore the oral combination appeared to be marginally less toxic than the i.p. route (Fig. 16B).

At a dose of 20mg/kg of inactivator in combination with temozolomide in DBA₂ mice, B.4205 and B.4280 were shown to be less acutely toxic than BeG, with B.4280 being less acutely toxic than B.4205 (Fig. 17).

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Figures 18 and 19 show that B.4280 (PaTrin-2) (i.p. at 20 mg/kg and p.o at 30mg/kg respectively) depletes ATase in human melanoma xenografts more completely and for a more extensive period than it does in host tissues.

Figure 20 show that despite the considerably higher initial level of ATase activity in the human breast tumour, B.4280 depletes ATase therein more completely and for a longer period of time than in murine host tissues. In this study using 30mg/kg B.4280 i.p. extensive depletion was seen in brain tissue, indicating the ability to cross the blood-brain barrier.

Figure 21 likewise shows that despite the considerably higher initial level of ATase activity in the human prostate tumour, B.4280 depletes ATase therein more completely and for a longer period of time than in murine host tissues. In this study using 20mg/kg

B.4280 i.p. relatively little depletion was seen in brain tissue, indicating by reference to Figure 20 that the ability of B.4280 to cross the blood-brain barrier may be dose-dependent.

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Figure 22 shows that B.4280 (20mg/kg i.p.) considerably increased the sensitivity of the human breast tumour xenograft to the growth inhibitory effects of temozolomide using a 5 day dosing schedule. This sensitization occurred despite the very high level of ATase in this tumour.

Figure 23 shows that a single dose of B.4280 (30mg/kg p.o.) considerably increased the sensitivity of the human melanoma tumour xenograft to the growth inhibitory effects of a single dose of the chloroethylating agent, fotemustine, without any substantial effect on toxicity.

Synthesis of Q^6 -(methylene[3 H])-(4-bromothenyl)guanine

Bromothenylaldehyde (0.79mg, 66.8 umoles was reacted with NaB[3 H) $_4$ (0.0167 mmoles, 60Ci/mmole) in isopropanol (350µl) for 1h at room temperature. The resulting [3 H]-4-bromothenylalcohol was extracted into pentane, dried, weighed and reacted with NaH (5.44mg), and the quaternary ammonium salt of guanine (15.55mg) in DMSO (250µl) for 1 hour at room temperature. The product was recovered by precipitation from acetic acid-ether (15µl glacial acetic in 1.5ml ether), washed with ether, dried and triturated with H $_2$ O. After washing with water, the final product was dried to constant weight. Figure 28 shows the scheme for synthesis of the radio-labelled B.4280.

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High performance liquid chromatography analysis

An aliquot of the product was dissolved in buffer A (10mM KH_2PO_4 containing 7.5% acetonitrile) and subjected to high performance liquid chromatography on an ODS-5 column. Elution at 1ml/min was with a linear gradient over 20 minutes from 100% A to 20% A:80% B (10mM KH_2PO_4 containing 80% acetonitrile). The effluent was monitored for UV absorption at 254nm and fractions (1

WO 97/20843 PCT/IE96/00084

- 41 -

min) were collected and assayed for radioactivity after addition of 10ml of Ecoscint A. It was shown that 96% of the radio activity co-chromatographed with authentic B.4280 (Figure 29).

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Incubation of an aliquot of the product with known amounts of pure recombinant human ATase resulted in the transfer of radioactivity to the protein (Figure 30), strongly suggesting that the mechanism of ATase inactivation involves the transfer of the thenyl group to the active site cysteine residue in the ATase molecule. Measurement of the amount of radioactivity transferred to protein indicated that the 0^6 -(0^8 H]-4-bromothenyl)guanine had a radiochemical purity of >96% and a specific activity of 16Ci/mmole.

15 <u>0</u>⁶([³H]-4-bromothenyl)guanine can be used as an alternative to the standard method, which presently uses [³H]-labelled substrate DNA, to determine the amounts of ATase, for example, in cell or tissue extracts. It may also be used to locate active ATase molecules in tumour and other tissue sections by incubation with 20 such sections on microscope slides followed by washing, autoradiography and histological staining. It may also be used to monitor the formation of the [³H]-labelled products of breakdown or metabolism of the agent after administration to mammals. It may also be used to determine the distribution of the B.4280 or its breakdown products in animal tissues and tumours by means of whole body autoradiography.

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Typical synthetic procedures

Type 1A.

O⁶-(4-Bromothenyl)hypoxanthine, B. 4292

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4-Bromothenyl alcohol (1.16 g, 6 mmol) was added to sodium hydride (60% in oil; 0.16 g, 2 mmol) and DMSO (1 ml). The solution was stirred for 30 min. The trimethylammonium salt (0.427 g, 2 mmol) was then added and stirring continued for 2.5 h at 20°C. The solution was cooled in an ice bath and poured into ether (60 ml) containing acetic acid (0.32 ml). A white precipitate was collected, triturated with water (4 ml) and collected again to give B. 4292 (436 mg, 69%) recrystallised from methanol.

Type 1B.

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06-Thenyl-2-methylhypoxanthine, B. 4350

DABCO salt from 6-chloro-2-methylpurine:

6-chloro-2-methylpurine (0.5 g, 3 mmol) was dissolved in a mixture of DMF (5 ml) and diglyme (25 ml). DABCO (0.66 g, 6 mmol) was then added. The mixture was stirred for 1 h and the precipitate collected to give the quaternary salt (700 mg, 82%). NMR (300 MHz, DMSO-d₆): shift in ppm 2.65 (s), 3.27 (t, J=7.5 Hz), 3.78 (s), 4.14 (t, J=7.5 Hz), 8.21 (s).

Thenyl alcohol (684 mg, 6 mmol) was added to sodium hydride (60% in oil; 80 mg, 2 mmol) and DMSO (0.5 ml). The solution was stirred for 30 min. The DABCO salt was then added and stirring continued for 5 h. The solution was then poured into ether (30 ml) containing acetic acid (0.15 ml). A precipitate was collected, triturated with water (4 ml) and collected again to give O6-Thenyl-2-methylhypoxanthine (96 mg, 35%) recrystallised from acetonitrile.

- 43 -

Type 1C.

06-(4-Bromothenyl)-2-fluorohypoxanthine, B. 4353

To 3.6 ml of 40% fluoroboric acid precooled to -25°C in a bath was added O⁶-(4-bromothenyl) guanine (326 mg, 1 mmol) with vigorous stirring. A solution of sodium nitrite (0.116 g, 1.7 mmol) in water (0.15 ml) was added dropwise over a period of 10 min. After 20 min, the solution was poured into ice. The mixture was then allowed to stand at 0°C for 15 h, then collected and dried to afford almost pure (t.l.c.) B. 4353 (180 mg, 55%). Flash chromatography (Hexane - Ethyl Acetate decreasing polarity little by little) afforded B. 4353.

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Typical synthetic procedures (continued)

Type 3D

O⁴-Thenyl-5-deazapterin, B. 4376

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a) N²-Pivaloyl-5-deazapterin

A mixture of 5-deazapterin^{33,34} (2.0g, 13.36mmol), 4-dimethylaminopyridine (0.22g, 1.8mmol) and pivalic anhydride (12ml) was heated to 165° C. Excess pivalic anhydride was distilled off and the residue dissolved in dichloromethane and applied to a pad of silica gel, and eluted with 2% methanol in dichloromethane. Evaporation and recrystallisation of the product from ethanol gave shiny cream coloured crystals (2.25g, 74%) of the pivaloyl derivative, m.p. 258-259°C; λ_{max} (MeOH) 277 nm; NMR (300MHz, DMSO-d₆) δ 1.28(s), 7.44(q), 8.43(dd), 8.88(dd), 11.4(s), 12.31(s).

b) N²-pivaloyl-O⁴-thenyl-5-deazapterin:

A suspension of N²-pivaloyl-5-deazapterin (0.492g, 2mmol) in tetrahydrofuran (8ml) was stirred for 10 min, and tri-n-butylphosphine (0.606g, 3mmol), thenyl alcohol (0.432g, 3mmol) and diisopropylazodicarboxylate (0.606g, 3mmol) were added successively. The reaction was allowed to proceed for 2h at room temperature and evaporation then gave an oil. Hexane was added to induce crystallisation. Filtration and recrystallisation from hexane gave bright yellow crystals of the thenyl derivative (0.32g, 47%) m.p. 107-108°C; λ_{max} (MeOH) 272, 311 nm; NMR (300MHz, DMSO-d₆) δ 1.28(s), 5.86(s), 6.98(q), 7.28(dd), 7.43(dd), 7.52(q), 8.46(dd), 8.89(dd).

c) B. 4376

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 N^2 -pivaloyl- O^4 -thenyl-5-deazapterin (0.28g, 0.82mmol) was heated for 24h under reflux with aqueous NaOH (3M, 2ml) and ethanol (1ml). The solvent was removed by evaporation and the residual solid dissolved in water. Acidification with acetic acid gave a white precipitate. Filtration and recrystallisation of the solid from ethanol gave white crystals of O^4 -thenyl-5-deazapterin (B. 4376), (0.107g, 51%).

Type 4D

O⁶-(4-Bromothenyl)-5-nitrocytosine, B.4380

Sodium hydride (60% in oil; 80mg, 2mmol) was added to a stirred solution of 4-bromothenyl alcohol (290mg, 1.5mmol) in dry DMSO (1ml). After 30 min, 4-amino-2-chloro-5-nitropyrimidine³⁵ (174mg, 1mmol) was added and the mixture heated at 50°C for 2h. The DMSO was removed in vacuo and the pH adjusted to 7 with aqueous acetic acid. After extraction into ethyl acetate, the product B. 4380 was crystallised from methanol (51mg, 15%).

Type 5

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S⁶-(4-bromothenyl)-6-thioguanine, B.4352

Sodium hydride (60% in oil; 44mg, 1.1mmol) was added to a stirred solution of 4-bromothenyl mercaptan (418mg, 2mmol) in dry DMSO (0.5ml). After 30 min, 2-amino-N,N,N-trimethyl-1H-purin-6-aminium chloride (228mg, 1mmol) was added and stirring continued for 1h. Acetic acid (0.12ml) and ether (30ml) were added and after decantation and trituration with fresh ether, B.4352 (38mg, 11%) was filtered off.

9-Substituted O⁶-(4-bromothenyl)guanines:

10 O⁶-(4-Bromothenyl)-9-(ethoxymethyl)guanine, B.4369

 O^6 -(4-Bromothenyl)guanine (652mg, 2mmol) was dissolved in sodium ethoxide (1M; 2ml, 2mmol). After 10 min, the ethanol was removed and the residue was dissolved in dry DMF. Chloromethyl ethyl ether (189mg, 2mmol) was added dropwise to the stirred solution under an atmosphere of argon. After 45 min, the solvent was removed. The oily product was crystallised from ethanol giving B.4369 (158mg) as needles. A further 118mg was obtained by flash chromatography of the mother liquor on silica gel with 5% ethanol in CH_2Cl_2 . Total yield, 39%.

O⁶-(4-Bromothenyl)-9-(2-hydroxyethoxymethyl)guanine, B. 4335

A stirred mixture of O⁶-(4-bromothenyl)guanine (294mg, 1mmol), (NH₄)₂SO₄ 20 (47mg) and hexamethyldisilazane (5ml) was heated at reflux for 3h. Volatile material was then evaporated under vacuum. The residue was stirred with benzene (15ml) and Hg(CN)₂ (344mg, 1.3mmol) under reflux for 30 min. A solution of (2acetoxyethoxy)methyl bromide {Ref 4 p33} (197mg, 1mmol) in benzene (10ml) was added, reflux maintained for 2h, and the cloudy diluted with chloroform (150ml). The organic phase was washed with saturated aqueous NaHCO3 (30ml), followed by KI 25 (1M; 30ml), dried over MgSO₄ and evaporated to give an oil (313mg). This oil was chromatographed on a silica gel column with CHCl3-MeOH (12:1) as eluant, yielding almost pure (t.l.c.) O-acetate (141mg) of B. 4335. Methanol (60ml) was saturated with dry ammonia and poured onto this Q-acetate in a flask which was tightly stoppered. After dissolution, stirring was stopped and the flask left closed overnight. Evaporation of methanol gave B. 4335 (135mg, 46%), 30 recrystallised from isopropanol.

O⁶-4-bromothenyl-9-(β-D-ribofuranosyl)guanine, B.4363

A mixture of 2',3',5'-tri-(O-acetyl)guanosine³⁶ (409mg, 1mmol), tri-n-butylphosphine (303mg, 1.5mmol) and 4-bromothenyl alcohol (290mg, 1.5mmol) in dry tetrahydrofuran (16ml) was stirred at room temperature for 45 min. Then diisopropyl azodicarboxylate (303mg, 1.5mmol) was added dropwise and the mixture stirred for 2h. The solution was evaporated leaving an oil which was dissolved in THF/MeOH/25% aqueous ammonia (1:1:1:5 ml) and kept for 48 h at 4°C.

WO 97/20843 PCT/IE96/00084

- 46 -

Adsorption on silica gel and column chromatography with CHCl₃/MeOH (15:1 to 10:1) gave the riboside B.4363 (205mg, 44%).

 O^6 -4-Bromothenyl-9-(β -D-2'-deoxyribofuranosyl)guanine, B.4379.

A mixture of 3',5'-di-(O-acetyl)-2'-deoxyguanosine³⁷ (554mg, 1.5mmol), tri-n-butylphosphine (666.6mg, 3.3mmol) and 4-bromothenyl alcohol (638mg, 3.3mmol) in dry tetrahydrofuran (40ml) was stirred at 80°C for 15 min. Then diisopropyl azodicarboxylate (666.6mg, 3.3mmol) was added dropwise and 15 min later, the reaction mixture was cooled and evaporated leaving an oil. This was dissolved in THF/MeOH/25% aqueous ammonia (1:1:1; 5 ml) and kept for 48 h at 4°C. Adsorption on silica gel and column chromatography with CHCl₃/MeOH (20:1) gave the 2'-deoxyriboside B.4379 (338mg, 51%).

9-(\beta-D-Arabinofuranosyl)-06-(4-bromothenyl)guanine, B.4368.

An alkoxide solution was made from sodium hydride (60% in oil; 60mg, 1.5mmol) and 4-bromothenyl alcohol (344mg, 1.8mmol) in dry DMSO (0.5ml) over 1 h. It was reacted with 2-amino-9-(β-D-arabinofuranosyl)-6-chloropurine³⁸ (151mg, 0.5mmol) and stirred for 5 min at room temperature, then 15 min at 60-65°C. Cooling and trituration with ether (50ml) and filtration yielded a solid which was dissolved in water (5ml), neutralised with acetic acid and treated with silica gel. Column chromatography with ethyl acetate/MeOH (19:1) gave the arabinoside B.4368 (87mg, 38%), pure on t.l.c.

O⁶-substituted guanines

These were made by the standard procedure from the quaternary salt 2-amino-N,N,N-trimethyl-1H-purin-6-aminium chloride and the appropriate alkoxide derived from the alcohol and sodium hydride in DMSO (cf. pp. 16d, 17, 18, 47 of 7/12/95).

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Compound, Test No.	O'-Substituent		Solvent for	Mr (docomn)	1 5.0		-			
	RCH,	%	Recrystn.	P.C. (decomb.)	rormuia	Molecular		(Analysis	
Type 1A. Hypoxanthines						WEIKIII.		- 1		اء
B, 4293	furfunyl	09	МеОН	154	CloHgN4O2	216	·			
B. 4291	thenyi	99	МеОН	168	C ₁₀ H ₈ N ₄ OS	232	Found	\$1.85	3.40	24.12
B. 4292	4-bromothenyl	69	МеОН	170	CloH,BrN,OS	311	Reg. Found	38.33	3.47	24.12 17.66
								0.00	07.7	18.00
1B. 2-Methylhypoxanthines B. 4347	benzyl	43	MeCN	191-193*	C ₁₃ H ₁₂ N ₄ O	240	Found	65.05	4.91	23.30
B. 4350	thenyl	35	MeCN	176-178*	C11H10N,OS	325	Req Found	53.63	3.90	23.32
IC. 2-Fluorohypoxanthines B.4353	4-bromothenyl	55	Column	142	O.H.BrFN OS	230	100	20.04	5	27.73
1D. 9-(2-Hydroxyethoxy-					Cloricality (CO	329				
methyl/guanines B. 4334	benzvl	46	HOrd.	140-1430						
		}		761-061	CISHTINSO)	315	Found		_	21.93
B. 4335	4-bromothenyl	42	i-PrOH	156-158	C ₁₃ H ₁₄ BrN ₅ O ₃ S	400	7			22.21 17.20
1E. 8-Hydroxyguanines	1	56			CloHeBiNsO2S.1/1 H2O	351	Found	34.53	3.53	19.50
6464.0	4-promotnenyi		Ag. EtOH	> 230			Req.	34.20	2.58	19.94
Type 24 & draming										
B.4270	4-fluorobenzyl	40	Аф. МсОН	> 280	C11H,FN,O	260	Found	\$1.50	3.85	29.44
B.4314	4-chlorothenyt	26	Ад. МсОН	> 200	C.H.CIN.OS	282.7	Req.			32.30
D 4300	-							38.24		29.73
D.4703	4-oromotnenyi	17	MeCN	> 190	C,H,BrN,OS	327	_		_	24 60

- 48 -

Compound, Test No.	O*-Substituent RCH2	Yicld %	Solvent for Recrystn.	M.p. (decomp.)	Formula	Molecular Weight		່ ວ	Analysis H	z
							Rcq.	33.04	2.16	25.69
1B. 8-Aza-7-deazaguanines B. 4310	benzyl		МсОН	160	C12H11N5O.H2O	259	Found	55.53	6.4	26.41
							Req.	55.59		27.02
B. 4340	4-fluorobenzyl	65	EtOH	188	C12H1cFN50.12H20	263.7	Found	53.82		26.97
2770	4 011010101111	5	הטם	24.3.24.40		200	Red	54.6		26.55
B. 4339	4-cniorocenzyi	76		h+7-7+7	7,7H,0 VEIOH	067	Red	50.15	2.87	23.43
B. 4343	piperonyl	50	ЕЮН	186			İ			5
B. 4348	furfuryl		ЕгОН	150°	C ₁₃ H ₁₁ N ₅ O ₃	285	Found	54.52		24.50
B. 4338	thenyl	89	EtOH	180	CloH,N,O2.14H,O	235.7	Req. Found	54.7 50.96	3.88	24.55
B. 4337	4-bromothenyl	79	ЕЮН	180	C _{lo} H ₉ N,OS	247	Req. Found	50.96 47.58		29.71
					CloHrBrN.OS	326				27.8
							Reg.		_	21.5
Type 3A. 8-Oxoguanines B. 4272	4-fluorobenzyl	41	Acctone	223-224	CuH,FN,O;	197				26.65
B. 4285	4-chlorobenzyl	63	Acctone	219-220	C ₁₁ H ₈ ClN ₅ O ₂	277.7	Req. Found			26.81
B. 4299	4-chlorothenyl	55	Acctone	164-165	C,H,CIN,O,S	283.7				25.22
B. 4287	4-bromothenyl	61	Acetone	170-172	C,H,BrN,O,S	328	Req. Found	33.30	2.13	24.69
3B. 8-Thloguanines B. 4296	benzyl	39	EIOH		C ₁₁ H ₉ N ₅ OS	259	†		 	
B.4286	4-fluorobenzyl	11	PLC	182-184	C ₁₁ H ₈ FN ₅ OS	277				······································

TABLE 1A (continued)

TABLE 1A (continued)

Compound, Test No.	O"-Substituent	Yield	Solvent for	M.p. (decomp.)	Formula	Molecular			Analysis	8
	NCH;	%	Kecrystn.	(<u>(</u> C)		Weight		ပ	Ħ	z
B. 4315	4-chlorothenyl	<u> </u>	МсОН		C,H,CIN,OS,	299.8	Found	36.27	2.04	23.07
B 4351	1	-	:				Req.	36.06	2.03	23.36
D: 4331	4-bromotneny:	-	MCH CH	156-160	C,H,BrN,OS,	344	Found	31.49	1.60	20.11
							Req.	31.41	1.76	20.35
3C. Pierins (O'-substituent) B. 4290	4-fluorobenzvi	*	HON	>210	0 20	17.		2000	6	
		:			OS21 1001 1450	7/7	round	79.70	5.00	20.07
B. 4316	4-chlorothenyl	4	MeOH	>170	C ₁₁ H ₈ CIN,OS	293.7	req. Found	44.93	2.84	23.72
2000	A here and 1	S	:0	90	:	(44.98	2.75	23.84
2071.7	4-010111011101191	G .	unaiwi	1/8-1/9	C11AgBIN ₅ OS	338		39.34	3.13	20.25
Type 4A. 2,4-diamino-6-hydroxypyrlmidines										
B. 4305	4-fluorobenzyl	98	C ₆ H ₆ /Petrol	133-134	C11H11FN4O	234		56.20	4.79	23.66
0 4304	4 -1.11.	-	:				Req.	56.40	4.73	23.92
D: +50	4-chiorocenzyi	7	CeHe	122-123	C'IIHIICIN'O	250.7		52.43	4.56	22.47
B. 4303	piperonyl	79	MeCN	168-171	C12H12N4O3	260	req.	55.31	4.42	22.35
B. 4307	thenyl	97	McPlı	100	SO. N. H. O.	333		55.38	4.65	21.52
									5. 4.	25.21
B. 4302	4-chlorothenyl	45	MePh	129-130	C ₉ H ₉ CIN ₄ OS	256.7	Found 6	 -	3.68	22.00
4B. 2,4-Diamino-6-hydroxy-5-							 	+-	3	6.17
B.4301	4-fluorobenzyl	92	NtcOH	>250	C11H10FN5O2	263 F	73	49.60		67.53
B. 4311	4-chlorothenyl	84	Acetone	>190	C,H,CIN,O,S	285.7	Req 3 Found - 3		3.83	26.61

TABLE 1A (continued)

Compound, Test No.	O*-Substituent	Yield	Solvent for	M.p. (decomp.)		Molocutor				
	RCH,	- 1	Recrystn.	(2)		Welght		ر پ 	Analysis C H N	
B. 4312	4-bromothenyl 62	62	Acctonc	200-201	C,H,BIN,O,S	330	Req.	m m i	2.82	24.51
4C. 24-Diaming-budgoon							Keq.	32.74	2.44	21.21
.c. sc. ammo-o-myaroxy-o-										
niropyrimidines						_		-		
B. 4308	piperonyl	29	DMF	>175	Civilino	105	Formar	47.44	- 50 7	- 6
•.										60.77
B. 4306	thenyl	34	MeOH	159-160	C.H.N.O.S	190	Req.	40.00 02 12.63	2.63	22.94
								10.77	7	77.77
						-	Red	40.44	3.39	26.21

Continuation of Table 1a

	Door	(°)		Weight		v	Analysis C H	× ×	
<u></u>	EIOH	215-216	C ₁₂ N ₁₀ N,OS	258					
									
15	МеОН	143-144	C,H,BrN,O,S	331					
69	СН,ОН	204-212	C ₁₃ H ₁₁ N ₅ O ₂ S	301	Found	50.25	3.60	23.66	
=	сн,он	180-184	CloHaBrNsOs. J CHsOH	342	Req. Found	51.82	3.68	23.24	
69		меон Сн,он Сн,он		143-144 204-212 180-184	143-144 C ₉ H,BrN ₄ O ₃ S 331 204-212 C ₁₅ H ₁₁ N ₅ O ₃ S 301 180-184 C ₁₀ H ₂ BrN ₅ O _{3, 2} CH ₃ OH 342	143-144 C ₉ H,BrN ₄ O ₃ S 331 204-212 C ₁₅ H ₁₁ N ₅ O ₃ S 301 180-184 C ₁₀ H ₂ BrN ₅ O _{3, 2} CH ₃ OH 342	143-144 C ₉ H,BrN ₄ O ₃ S 204-212 C ₁ H ₁₁ N ₃ O ₃ S 180-184 C ₁₀ H ₂ BrN ₃ O ₃ CH ₃ OH	143-144 C ₉ H,BrN ₂ O ₃ S 331 50.25 204-212 C ₁₃ H ₁₁ N ₃ O ₂ S 301 Found 50.25 180-184 C ₁₆ H ₂ BrN ₃ O _{3, 2} CH ₃ OH 342 Found 35.07 Req. 51.82	143-144 C ₉ H,BrN ₄ O ₃ S 331 204-212 C ₁₃ H ₁₁ N ₅ O ₂ S 301 Found 180-184 C ₁₀ H ₄ BrN ₅ O _{3, \(\begin{array}{c} \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \}

TABLE 1A (continued)

Compound, Test No.	9-Substituent Yield	Yield %	Solvent for Recrystn.	M.p	Formula	Molecular Wolch		Analysis ^d	
				(2)		Meigu	C	Ŧ	z
B.4369	ethoxymethyl	39	ЕтОН	134-5	134-5 C ₁₃ H ₁₄ BrN ₅ O ₂ S	384	40.58	3.71	17.97
B.4370	n-octyloxymethyl	39	ЕтОН	06	C ₁₉ H ₂₆ BrN ₅ O ₂ S	468	(40.64 48.97	3.67	18.23) 14.82
B.4334 ^b	2-hydroxy-	46	i-PrOH	150-2	C ₁₅ H ₁₇ N ₅ O ₃	315	(48.72 57.19 (57.13	5.59 5.43	14.95) 21.93 22.21)
B.4335	2-hydroxy-	42	i-PrOH	156-8	$C_{13}H_{14}BrN_5O_3S$	400	39.16	3.68	17.20
B.4363	β-D-ribo-	4			C ₁₅ H ₁₆ BrN ₅ O ₅ S	458			-
B.4368	furanosyl β-D-arabino- furanosyl	38			C ₁₅ H ₁₆ BrN ₅ O ₅ S	458			51 -
B4379	β-D-2-deoxyribo- 51	51			C ₁₅ H ₁₆ BrN ₅ O ₄ S	442			
* Found, with required values	nranosyl								

Found, with required values in parenthesis.

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O'-Substituent RCH2 Ams (McOH) Sulppm from TMS, (CD3)2SO, J(Hz)	5.60(s), 6.53(dd, 3.1, 1.9), 6.69(d, 3.1), 7.76(dd, 1.9, 0.9) 8.39(s), 8.55(s) 5.83(s), 7.08(dd, 5.1, 3.4), 7.35(d, 3.4), 7.6(d, 5.1), 8.39(s), 8.51(s) 5.80(s), 7.38(d,1.3), 7.73(d, 1.3), 8.42(s), 8.58(s)	2.61(s), 5.60(s), 7.50(m), 8.32(s) 2.63(s), 5.77(s), 7.05(dd, 5.1, 2.4, 7.33(d), 2.4), 7.58(dd, 5.1, 1.0) 8.26(s), 13.22(s)		5.77(s), 7.4(d, 1.5), 7.77(d, 1.5), 8.45(s), 13.64(bs).		3.48(m), 4.70(s), 5.45(s), 6.59(s), 7.45(m), 8.03(s), 3.49(m), 4.71(s), 5.45(s), 5.66(s), 6.65(s), 7.30(d, 1, 5), 7.27d, 1, 5), 8.04(s),	5.54(s), 6.24(s), 7.33(d, 1.4) 7.70(d, 1.4), 10.49(s) 11.12(s)	5.57(s), 7.04(s), 7.28(m), 7.65(m), 15.38(s). 5.71(s), 7.13(s), 7.41(d, 1.5), 7.66(d, 1.5), 15.42(s). 5.73(s), 7.12(s), 7.43(d, 1.5), 7.76(d, 1.5), 15.39(s).
1 _{2 λπαε} (ΜcOH) (nm)	252 240 251	256 240		233, 255		247, 283 3 245, 284 3	239, 293 5.	288 5. 288 5. 287 5.
O ^e -Substituent RCE	furfuryl thenyl 4-bromothenyl	benzyl thenyl		4-bromothenyl		benzyl 4-bromothenyl	4-bromothenyl	4-tluorobenzyl 4-chlorothenyl 4-bromothenyl
Compound Type, Test No.	Type 1A. Hypoxanthines B. 4293 B. 4291 B. 4292 Type 1B. 2-Methyllypoxanthines.	B. 4347 B. 4350	Type 1C. 2-Fluorohypoxanthines	B. 4353	Type 1D, 9-(2- Hydroxyethoxymethyl)guanines	B. 4334 B. 4335	Type 1E. 8-Hydroxygnanines B. 4349	Type 2A. &-Azaguanines B. 4270 B. 4314 B. 4289

TABLE 1B (continued)

	5.50(s), 6.68(s), 7.74(m), 7.82(s.), 12.87(hs)	5 49(s), 6 70(s), 7.20(m), 7 61(m), 7 82(s), 12 88(bs)	5.50(s), 6.69(s), 7.49(d. 8.4), 7.56(d. 8.4), 7.83(s) 12.90(s)	5.39(s), 6.05(s), 6.69(s), 6.94(d, 7.9) 7.04(dd, 7.9, 1.5), 7.14(d, 1.5), 7.80(s)	12.86(bs),	5.46(s), 6.52(s), 6.70(s), 6.71(s), 7.73(s), 7.79(s), 12.85(bs),	5.69(s), 6.73(s), 7.07(d, 3.5), 7.35(s), 7.60(d, 1.1), 7.79(s), 12.90(bs)	5.65(s), 6.76(s), 7.38(s), 7.72(d, 1.3), 7.81(s), 12.91(bs).		5.62(s), 7.30(t, 9.1), 7.68(m), 7.91(s), 7.97(s),	5.63(s), 7.53(d, 8.3), 7.65(d, 8.3), 7.90(s), 7.97(s).	5.78(s), 7.46(d, 1.6), 7.72(d, 1.6), 7.95(s), 8.01(s).	5 79(s), 7 49(d. 1 6), 7 81(d. 1 6), 7.95(s), 8 01(s)			5.59(s), 7.29(t 8 9), 7 51(hs), 7 67(m)	5.75(s), 7.44(d. 1.6), 7.55(bs), 7.69(d. 1.6)	5.78(s), 7.45(d, 1.6), 7.46(bs), 7.75(d, 1.6)		5 56(s), 7.29(t. 8.85), 7.44(bs), 7.66(m), 8 45(d. 1 8), 8.82(d, 1.8).	5.71(s), 7.41(d, 1.6), 7.47(bs), 7.67(d, 1.6), 8.46(d, 2.0), 8.83(d,2.0)	5.73(s), 7.44(d, 1.6), 7.50(bs), 7.77(d, 1.6) 8.46(d, 2) 8.83(d, 2).
	277	278	276	282		277	278	278		257, 341	256, 340	252, 343	253, 343		227, 361	235, 362	228, 360	228, 361		232, 264(sh). 362	364	
	benzyl	4-fluorobenzyl	4-chlorobenzył	piperonyl		furfunyl	thenyl	4-bromothenyl		4-fluorobenzył	4-chlorobenzył	4-chlorothenyl	4-bromothenyl		benzyl	4-fluorobenzyl	4-chlorothenyl	4-bromothenyl		4-Auorobenzyl	4-chlorothenyl	4-bromothenyl
Type 2B. 8-stan-7-degrammes	B. 4310	B. 4340	B, 4339	B 4343	•	B. 4348	B. 4338	B. 4337	Type 3A. &-(Dxagnanines	B. 4272	B. 4285	B. 4299	B. 4287	Type 3B. 8-Thinguenines	B. 4296	B. 4286	B. 4315	B. 4351	Type 3C. Pierins (O'-substituent)	B. 4290	B. 4316	B. 4288

5.33(s), 6.05(s), 6.95(d, 8.0), 7.00(dd, 8.0, 1.4), 7.10(d, 1.4); 7.26(bs), 7.3 7.96(bs).
5.59(s), 7.03(dd, 5.1, 3.5), 7.28(d, 3.5), 7.32(bs), 7.56(d, 5.1), 7.94(bs).

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B. 4306

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piperonyl

TABLE 18 (continued)

			•) + -		
	5.10(s), 5.19(s), 5.96(s), 6.10(s), 7.19(t, 8.8), 7.44(dd, 8.8, 5.8). 5.11(s), 5.22(s), 5.96(s), 6.10(s), 7.44(s). 5.09(s), 5.11(s), 5.97(s), 6.03(s), 6.07(s), 6.91(d, 1.1), 7.00(s). 5.08(s), 5.40(s), 6.00(s), 6.10(s), 7.03(dd, 8.1, 3.5)7.20(dd, 8.1, 1.1), 7.54	(dd, 3.5, 1.1). 5.08(s), 5.35(s), 6.03(s), 6 13(s), 7.19(s), 7.55(d 1.6).		5.59(s), 7.26(m), 7.65(m), 7.80(bs), 7.85(bs), 8.00(bs), 10.05(bs). 5.73(s), 7.40(d, 1.6), 7.66(d, 1.6), 7.94(s), 7.98(d, 2.7), 8.11(d, 4.2) 10.03	(d, 4.2). 5.75(s), 7.42(d, 1.4), 7.75(d, 1.4), 7.93(s), 7.98(s), 8.12(d, 4.0), 10.04 (d, 4.0)	5.33(s), 6.05(s), 6.95(d. 8.0), 7.00(dd. 8.0, 1.4), 7.10(d. 1.4), 7.26(h.), 7.2
	238, 267 238, 268 236, 267 235, 267	236, 265		336 335	335	288,330
	4-fluorobenzyl 4-chlorobenzyl piperonyl thenyl	4-chlorothenyl		4-fluorobenzyl 4-chlorothenyi	4-bromothenyl	piperonyl
Type 4A. 2,4-diamino-6-hydroxy-pyrimidines	B. 4305 B. 4304 B. 4307	B. 4302	Type 4B. 2,4-Diamino-6- hydroxy-5-nitraxapyrmidines	B. 4301 B. 4311	B. 4312	Type 4C. 2,4-diamino-6-hydraxy- 5-nitropyrimidines B. 4308

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Compound Type, Test No.	Substituent RCH,	λ _{max} (MeOH) (nm)	δ _H [ppm from TMS, (CD ₃) ₂ SO,]J(Hz)
Type 3D 5-Deazapterins (04-substituent)			
B.4376	thenyl	248, 309	5.54(s), 6.96(q), 7.716(dd), 7.38(dd), 7.41(q), 8.39(dd), 8.79(dd).
Type 4D 5-Nitrocytosines (C'-substituent) B.4380	4-bromothenyl	255 sh, 334	5.19(s), 7.20(s), 7.56(d), 8.24(s), 8.70(s), 8.90(s).
Lype 5 0-1 mognanines (3 -substituent) B.4228	piperonyl	245, 311	4.56(s), 6.06(s), 6.55(s), 7.03(d), 7.06(d), 7.14(s),
B.4352	4-bromothenyl	241, 314	4.77(s), 6.52(s), 7.18(d), 7.51(d), 7.93(s), 12.61(b

TABLE 1B (continued)

δ _H [ppm from TMS, (CD ₃) ₂ SO,]J(Hz)		3.48(m), 4.70(s), 5.45(s), 6.59(s), 7.45(s), 8.03(s).		7.72(a, 1.5), 8.40(s). 3.54(m), 3.63(m), 3.91(dd), 4.12(dd), 4.48 (ddd), 5.12(dd), 5.18(d), 5.45(d), 5.66(s), 5.80(dd), 6.61(s), 7.38(d), 7.71(d),	8.15(s). 3.64(m), 3.76(dd), 4.07(m), 5.09(dd), 5.51(d), 5.53(m),	0.13(d), 0.00(d), 7.37(d), 7.71(d), 7.95(s) 2.39(ddd), 2.72(ddd), 3.65(ddd), 3.98(dd), 4.40(dd), 5.11(s) 5.41(d), 5.80(s), 6.38(dd), 6.67(s), 7.49(d), 7.83(d), 8.25(s).
λ _{max} (MeOH)	245, 284 245, 284 245, 284	245, 283	245, 284	•	245, 284	
9-Substituent Yield %	ethoxymethyl n-octyloxymethyl	2-hydroxy-	2-hydroxy-	eutoxymetnyt β-D-ribo- furanosyl	β-D-arabino-	tutanosyt β-D-2-deoxyribo- furanosyl
Compound Type, Test No.	B.4369 B.4370	B.4334 a	B.4335	B.4363	B.4368	B.4379

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	TABLE 2 INACTIVATOR TYPE	Ι ₅₀ (μΜ) hAT	T 1/2 (h) in PBS
5	1A		
	B.4291 0^6 -(thenyl)-hypoxanthine	1.9	>20
	B.4293 $\underline{0}^{6}$ -(furfuryl)-hypoxanthine	28	>16
10	B.4292 $\underline{0}^{6}$ -(4-bromothenyl)-hypoxanthine	0.3	>16
	0^6 -(benzyl)-hypoxanthine b	85	
	1B		
15	B.4347 0^6 -(benzyl)-2-methylhypoxanthine	75	
	B.4350 $\underline{0}^{6}$ -(thenyl)-2-methylhypoxanthine	14	
	1C		
20	B.4353 0^6 -(4-bromothenyl)-2-fluorohypoxanthine	1.4	
	0^6 -(benzyl)-2-fluorohypoxanthine a	48	
	10		
25	B.4334 0^6 -(benzyl)-9-(2-hydroxyethoxymethyl) guanine	8	>20
	B.4335 <u>O</u> ⁶ -(4-bromothenyl)-9-(2-hydroxy ethoxymethyl)guanine	See Table	: 3
30	1E		
	B.4349 $\underline{0}^{6}$ -(4-bromothenyl)-8-hydroxyguanine	See Table	3
	<u>0</u> 6-(benzyl)-8-hydroxyguanine ^a	0.3	
35	2A		
	B.4270 <u>O</u> ⁶ -(4-fluorobenzyl)-8-azaguanine	0.08	

	TABLE 2 (continued) INACTIVATOR TYPE	1 ₅₀ (μ M) hAT	T 1/2 (h) in PBS
5	B.4314 $\underline{0}^{6}$ -(4-chlorothenyl)-8-azaguanine	See Table 3	
	B.4289 $\underline{0}^{6}$ -(4-bromothenyl)-8-azaguanine	0.045	>10
10	0^6 -(benzyl)-8-azaguanine a	0.07	
	B.4310 $\underline{0}^6$ -(benzyl)-7-deaza-8-azaguanine	0.01	>16
15	B.4340 <u>O</u> ⁶ -(4-fluorobenzyl)-8-aza-7-deazaguanine	0.018	>16
••	B.4339 <u>O⁶-(4-chlorobenzyl)-8-aza-7-deazaguanine</u>	0.02	1.5
	B.4343 <u>O⁶-(piperonyl)-8-aza-7-deazaguanine</u>	See Table 3	
20	B.4348 <u>O⁶-(furfuryl)-8-aza-7-deazaguanine</u>	0.036	0.27
	B.4338 $\underline{0}^{6}$ -(thenyl)-8-aza-7-deazaguanine	0.01	
25	B.4337 <u>O</u> 6-(4-bromothenyl)-8-aza-7-deazaguanine	0.007	>20
23	3A		
	B.4272 <u>O⁶-(4-fluorobenzyl)-8-oxaguanine</u>	See Table 3	
30	B.4285 <u>O</u> ⁶ -(4-chlorobenzyl)-8-oxaguanine	0.225	4.6
	B.4299 0^6 (4-chlorothenyl)-8-oxaguanine	0.243	9.2
	B.4287 $\underline{0}^{6}$ -(-4-bromothenyl)-8-oxaguanine	0.24	2.6
35	B.4232 <u>O</u> ⁶ -(benzyl)-8-oxaguanine	0.25	

	TABLE 2 (continued) INACTIVATOR TYPE	Ι ₅₀ (μ Μ) hAT	T 1/2 (h)
	3B	IIAI	111 7 113
5	B.4296 $\underline{0}^{6}$ -(benzyl)-8-thiaguanine	0.02	>17
	B.4286 0^6 -(4-fluorobenzyl)-8-thiaguanine	0.03	>17
10	B.4315 0^6 -(4-chlorothenyl)-8-thiaguanine ^C	0.006	
	B.4351 $\underline{0}^{6}$ -(4-bromothenyl)-8-thiaguanine	See Table 3	
	3C	•	,
15	B.4290 <u>0</u> 4-(4-fluorobenzyl)-pterin	0.088	>10
	B.4316 $\underline{0}^4$ -(4-chlorothenyl)-pterin	See Table 3	
	B.4288 $\underline{0}^4$ -(4-bromothenyl)-pterin	0.025	>10
20	4A		
	B.4305 2,4-diamino-6-(4-fluorobenzyloxy)pyrimidine	4.0	>16
25	B.4304 2,4-diamino-6-(4-chlorobenzyloxy)pyrimidine	5.0	>16
23	B.4303 2,4-diamino-6-(3,4-piperonyloxy)pyrimidine	0.8	12.5
	B.4307 2,4-diamino-6-(thenyloxy)pyrimidine	0.4	4.2
30	B.4302 2,4-diamino-6-(4-chlorothenyloxy)pyrimidine	0.17	>16
	2,4-diamino-6-(benzyloxy)pyrimidine ^a	15	•
	4B		
35	B.4301 2,4-diamino-6-(4-fluorobenzyloxy)-5- nitrosopyrimidine	0.0175	>16

	TABLE 2 (continued) INACTIVATOR TYPE	I ₅₀ (uN) hAT	T 1/2 (h) in PBS
5	B.4311 2,4-diamino-(4-chlorothenyloxy)-5- nitrosopyrimidine	See Table 3	
	B.4312 2,4-diamino-6-(4-bromothenyloxy)-5- nitrosopyrimidine	0.045	4
10	2,4-diamino-6-(benzyloxy)-5- nitrosopyrimidine ^a	0.06	
	4C		
15	B.4306 2,4-diamino-6-(thenyloxy)-5- nitropyrimidine	2.3	>16
13	B.4308 2,4-diamino-6-piperonyloxy-5-nitropyrimidine	0.5	9.2
	2,4-diamino-6-benzyloxy-5-nitropyrimidine ^a	0.06	
	4D		
20	B.4380 $\underline{0}^{2}(4-bromothenyl)-5-nitrocytosine$	50	
	5		
2 5	B.4228 \underline{S}^6 -(piperonyl)-6-thioguanine	50	
	B.4352 \underline{S}^{6} -(4-bromothenyl)-6-thioguanine	8	
	Comparative		
30	B.4376 $\underline{0}^{6}$ -thenyl-5-deazapterin	1,600	
	Results for some 9-substituted $\underline{0}^6$ (4-bromothenyl)guanines are included in Table 7.		
35	a Data taken from Chae et al, <u>J. Med. Chem.</u> 1 b Data taken from Moschel et al., <u>J. Med. Che</u> c B.4315 Raji I ₅₀ (uM) 0.002	995, 38, 359-3 <u>m.</u> 1992, <u>35</u> , 4	65 486-4491.
	Blank Space = not done.		

TABLE 3

		_				- 61 -	· 							
Raji cell toxicity at	10 µM 'B'	(% Growth)		111.21+23.3	113.0+31.0	55.5+7.3 (D ₅₀ 16 μM)	85.5+20.0	98.4+12.2	97.0+10.0	90.0+13.0	117.5+29.1	82.2+11.0	69.8+10.3 (D50 44μM)	
Solubility in Water	(mg/m])			0.002	0	3.46 Not done	0.3	0.009	0.01	0.002	Not done	0.023	Not done	
)r	Œ	рМ)	0.1		1.4	3.46	1.4	1.0	1.6	2.4	1.2	1.03	5.5	
n facto "B")	ОМП	ation (0.5		:		;	;	:	:	:	:	&	
itisatio ol / D,	10201	ncentr	1.0	:	8.25	4.61	13.2	3.5	2.12	33	1.32	1.89	33	
Raji cell sensitisation factor (D ₃₀ control / D ₅₀ 'B')	TEMOZOLOMIDE	Inactivator concentration (µM)	10	1.41	73.3	84	99	38	9.5	50.8	18.1	27.5	09	
	BCNU	Inactiv	10	1.88	8.0	7.62	6.4	5.33	3.81	4.8	4.8	4.33	0.9	
T ^{1,2} (h)	Assay			2.6	12.5	>48	32	>48	3	>48	12	>75	>48	
T ^{1,2} (h)	PBS			5.7	10	>19	>19	>19	7.5	7.3	>16	>64	>16	
I _{so} ada	(M _I)			>1000	>1000	>1000	>1000	>1000	>1000	>1000	>1000	>1000	>1000	
	(hMq)			>1000	1.8	2	3.8	156	30	0.08	5.8	17	0.85	
I _{so} chAT	(hM)			0.04	0.02	0.03	0.04	1.8	0.02	0.02	0.03	0.01	0.03	
l _{so} rAT	(hMq)			0.075	0.016	0.037	0.03	6.5	0.045	0.074	0.027	0.076	0.019	
I _{so} mAT	(m Mil)	-		0.125	0.008	0.073	0.068	15.63	0.31	0.043	0.071	0.2	0.05	
I _{so} Raji (µM)				0.023	0.009	0.012	0.011	0.02	0.0085	0.007	0.005	0.1	0.003	
I _{so} hAT	(M ^{rl})			0.05	0.009	0.011. 0.012	0.025	0.33	0.007	0.018	0.003	0.04	0.003	
Mol				261	286	283	294	400	285	342	344	241	326	
Inactivator				B4272	B4311	B4314	B4316	B4335	B4343	B4349	B4351	BeG	PaTrin-2 B.4280	

--- = Not Done

- 62 -

TABLE 4

EFFECT OF INACTIVATOR PRETREATMENT ON SENSITISATION OF VAROUS HUMAN CANCER CELL LINES TO TEMOZOLOMIDE

INACTIVATOR	S	SENSITIS	ATION FACT	OR (D _{so}	control/D	so 'B')	
	MCF-7	PC3	DU145**		R	AJI	
	Inac	tivator do (10 µM)	se	10	Inactivate	or dose (₁ 0.5	μ M) 0.1
B4311		5.56	3.75	73.3	8.25		1.4
B4314*		2.0	1.71	84.0	4.61		3.46
B4316	8.0	7.6	3.53	66	13.2		1.4
B4349	4.8	3.6	4.0	50.8	33.0		2.4
BeG	2.94	2.88	5.45	27.5	1.89		1.03
PaTrin-2	3.13	4.6	4.14	60	33.0	8.0	5.5

^{*} Toxic to Raji cells at $10 \,\mu\text{M}$ ** Sensitisation factor = D_{60} control/ D_{60} 'B'

⁻⁻⁻ Not done

TABLE 5

EFFECT OF INACTIVATOR PRETREATMENT ON SENSITISATION OF VARIOUS HUMAN CANCER CELL LINES TO BCNU

INACTIVATOR (10 µM)	SENS	ITISATION	N FACTOR	(D _{so} cont	10ľD∞,	В')
	MCF-7	PC3	DU145**		RAJI	
				Inactiva 10	ator dose 1.0	e (μM) 0.1
B4311		1.47	1.56	8.0		
B4314*		1.46	1.25	7.62	7.6	3.45
B4316	1.37	1.35	3.57	6.4		
B4349	1.85	1.63	2.78	4.8		
BeG	1.94	1.41	1.79	4.33		
PaTrin-2	1.61	2.11	2.08	6.0		·

^{*} Toxic to Raji cells at 10 µM

** Sensitisation factor = D₆₀ control/D₆₀ 'B'

⁻⁻⁻ Not done

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Test No.	Test No. O ⁶ -Substituent	Yield % (based on solvate)	Solvent for recrystn.	M.p. (decomp.) (°C)	Formula		∢	Analysis		
7007	4 L	i	•						z	
5.4280	4-bromothenyl	73	MeOH	204-205	C ₁₀ H ₈ BrN,OS				21.46	
, מלץ	91	;	i			Req.	36.82	2.47	21.47	
1874.0	o-chlorothenyl	39	MeCN	155-158	C ₁₀ H ₈ CIN ₅ OS	Found			24.10	
ָר מ	4.		•			Req.			24.86	
5.4783	5-cyanothenyl	01	МеОН	200 upwards	CIIHINOS.	Found			28.24	
B.4294	5-methylenloh-				0.5 H ₂ O	Req.			29.88	
	inylthenyl	32	MeOH	200 upwards	C11H11N5O2S2	Found	42.58	3.62	22.27	
6	•	•				Reg	42.71	3.58	22.64	
B.4298	4-chlorothenyl	34	MeCN	194-198	C ₁₀ H ₈ CIN,OS	Found	42.70	2.94	24.84	
0000		;	•			Req	42.63	2.86	24.86	•
0.4300	4-methoxythenyl	44	МеОН	189-190	C11H11N5O2S	Found	47.73	4.15	25.05	' -
4313		Š				Req	47.64	4.00	25.26	_
5.4513	5-0romo-3-	9.7	MeCN	190 upwards	C ₁₀ H ₈ BrN,OS	Found	37.02	2.43	20.95	
0 4317	(menyimetny)	;	;			Req	36.82	2.47	21.47	
D.451/	4-cyanotnenyi	32	MeOH	213-216	C11H ₈ N ₈ OS	Found	48.50	2.84	30.66	
D 4210	4 6 13-11-11-14 A	ç				Req	48.52	2.96	30.87	
5.43.18	4,5-dichlorothenyl	38	MeOH	210 upwards	C ₁₀ H ₂ Cl ₂ N ₅ OS.	Found	35.94	2.67	20.96	
	Called A	9			14,0	Req	35.94	2.71	20.96	
D.4321	z-cnioro-4-picolyi	0	МеОН	234 upwards	C ₁₁ H,CIN,O	Found	47.15	3.52	29.32	
2007	,	(Req	47.75	3.29	30.37	
0.4330	5-promonurury	39	MeOH	180 upwards	C10HBIN,O2	Found	38.22	2.71	21.93	
					0.25 H ₂ O	Red	38.18	2.72	22.26	

6.6 mmol alcohol per mmol quaternary salt used in synthesis. Dimethylformamide reaction solvent.

O'-Substituted guanines (continuation of Table 6a,)

	•		`							
Compound, Test No.	O ⁶ -Substituted RCH ₂	Yield %	Solvent for Recrystn.	M.p. (decomp)	Formula	Molecular Weight		Analysis		
				<u>(</u>			ပ	Ξ.		Z
B.4282	3-picolyl N-oxide	54	МеОН	244-254	C11H10N6O2	258				
B.4309	5-methylsulphonyl- thenyl	12	ЕtОН	206-209	C ₁₁ H ₁₁ N ₅ O ₃ S ₂ .½ C ₂ H ₅ OH	348	Found Req.	41.46	3.83	20.13 20.10
B.4319	6-chloro-3-picolyl	58	МеОН	> 215	C1H,CI N,O	285.7	Found Req.	46.01 46.25	3.49	29.05 29.42
B.4320	5-bromo-3-picolyl	2 6	МеОН	> 220	C11H9BrN6O .½ H2O	330	Found Req.	40.02 39.87	3.05	25.28 25.28
B.4354	4-isothiazolyl	28	МеОН	> 200	C,H ₈ N,OS .½ H ₂ O	261.8	Found Reg.	41.59	3.64	31.53 32.16
B.4356 B.4357	4-methylthiothenyl 5-iodo-3-thienyl- methyl	30 23	МеОН МеОН	> 200	C11H11N5OS2 C10H8I N5OS	293.4 373	•			
B.4361	4-methyl- sulphonylthenyl	95	МеОН	170-172	$C_{11}H_{11}N_5O_3S_2$	325	Found Req.	40.2	3.39	21.01
B.4366	naphtho[2,1-b]- thiophen-2-yl- methyl	81	МеОН	> 150	C ₁₈ H ₁₃ N ₅ OS	347				
B.4373 B.4377	4-azidothenyl 4-methyl- sulphinylthenyl	37 55	МеОН МеОН	> 195° 204-206	C ₁₀ H ₈ N ₈ SO C ₁₁ H ₁₁ N ₅ O ₂ S ₂	288 309				
B.4378	5-phenylthenyl	54	CH,CN	>170	C ₁₆ H ₁₃ N ₅ OS	323				

TARIF 6

δ _H [ppm from TMS; (CD ₃) ₂ SO], J (Hz)	5.65(s), 6.40(s), 7.37(d), 7.71(d), 7.85(s), 12.49(s).	5.59(s), 6.40(s), 7.06(d), 7.22(d), 7.87(s), 12.47(bs).	5.73(s), 6.46(s), 7.49(d), 7.87(s), 7.92(d), 12.54(bs).	2.93(s), 5.73(s), 6.41(s), 7.40(d), 7.52(d), 7.88(s), 12.52(bs).	5.64(s), 6.42(s), 7.34(d), 7.62(d), 7.86(s), 12.51(s).	3.75(s), 5.57(s), 6.37(s), 6.60(d), 7.01(d), 7.85(s), 12.48(s).	5.42(s), 6.38(s), 7.40(d), 7.72(d), 7.85(s), 12.47(s).	5.68(s), 6.44(s), 7.74(d), 7.86(s), 8.60(d), 12.50(s).	5.58(s), 6.45(s), 7.41(s), 7.87(s), 12.52(s).	5.58(s), 6.36(s), 7.51(bs), 7.61(bs), 7.91(bs), 8.44(bs), 12.56(bs).	5.42(s), 6.39(s), 6.64(d), 6.78(d), 7.85(s), 12.49(s).
.Ն _{Պ&Հ} (nm)(MeOH)	238, 284 (RCH,OH:233)	247, 284 (RCH ₂ OH:245).	247, 272	243, 284(sh) [RCH ₂ OH:240, 274(sh)].	238, 284 (RCH ₂ OH:240)	245(sh), 282 (RCH ₂ OH: 258)	240, 284 (RCH ₂ OH: 236).	244, 284 (RCH ₂ OH: 244)	243, 285 (RCH ₂ 0H:243).	241, 272(sh), 285 [RCH ₂ OH: 262, 268(sh)].	220, 284 (RCH ₂ OH: 223)
O*-Substituent	4-bromothenyl	5-chlorothenyl	5-cyanothenyl	5-methylsulphinylthenyl	4-chlorothenyl	4-methoxythenyl	5-bromo-3-thienylmethyl	4-cyanothenyl	4,5-dichlorothenyl	2-chloro-4-picolyl	5-bromofurfuryl
Test No.	B. 4280	B. 4281	B. 4283	B. 4294	B. 4298	B. 4300	B. 4313	B. 4317	B. 4318	B. 4321	B. 4336

 $\underline{0}^6$ -Substituted guanines (continuation of Table 6B)

Compound Type, Test No.	O ⁶ -Substituent RCH ₂	λ _{mu} (McOH) (nm)	δ _{II} [ppm from TMS, (CD ₃) ₁ SO,]J(Hz)
B.4282	3-picolyl N-oxide	271	5.48(s), 6.41(s), 7.47(m), 7.87(s), 8.22(m), 8.42(s), 12.52(s).
B.4309	5-methylsulphonyl- thenyl	242, 284	5.75(s), 6.43(s), 7.47(d), 7.74(d), 7.87(s), 12.52(s).
B.4319	6-chloro-3-picolyl	242, 276	5.53(s), 6.38(s), 7.59(d), 7.87(s), 8.05(dd), 8.64(d), 12.48(s).
B.4320	5-bromo-3-picolyl	242, 281	5.53(s), 6.41(s), 7.86(s), 7.86(s), 8.26(dd), 8.73(d), 8.78(d), 12.50(s).
B.4354	4-isothiazolyl	244, 284	5.58(s), 6.41(s), 7.84(s), 8.81(s), 9.22(s), 12.47(s).
B.4356	4-methylthio-thenyl	236, 283	2.48(s), 5.62(s), 6.40(s), 7.26(m), 7.85(s), 12.48(s).
B.4357	5-iodo-3-	240, 283	5.43(s), 6.38(s), 7.48(d), 7.77(s), 7.84(s), 12.47(s).
	threnylmethyl		
B.4361	4-methylsulphonyl- thenyl	240, 285	3.26(s), 5.70(s), 6.40(s), 7.72(s), 7.85(s), 8.38(d), 12.49(s)
B.4366	naphtho[2, 1-b]-	244, 286sh	5-90(s), 6-47(s), 7-60(t), 7-69(t), 7-86(t), 8-04(dd),
B.4373	disciplinations 4-azidothenvi	227, 280	8.44(s), 8.31(d), 12.31(s). 5.64(s), 6.36(s), 7.20(s), 7.28(s), 7.84(s), 12.47(s)
B.4377	4-methylsulphinyl-	241, 285	2.82(s), 5.68(s), 6.33(s), 7.60(s), 7.82(s), 8.01(s), 12.45(s)
B.4378	5-phenylthenyl	244sh, 289	5.67(s), 6.32(s), 7.31(m), 7.41(m), 7.41(m), 7.63(d), 7.82(s), 12.43(s).

TABLE Z				
INACTIVATOR	M.E	^I 50 (µМ) . hAT	Raji I ₅₀ (µМ)	Stability T 1/2(h) By Spec
B.4280 <u>Q⁶-(4-bromotheny</u> l)guanine	326	0.0034		
B.4281 <u>Q⁶-(5-chlorotheny</u> l)guanine	281.7	0.004		>10
B.4282 <u>Q⁶</u> -(oxido-3-picoly1)guanine	276	1.4		>20
B.4283 <u>Q⁶</u> -(5-cyanothenyl)guanine	272	0.005		>20
B.4294 <u>O</u> ⁶ -(5-methylsulphinylthenyl)guanine	309	0.03		>10
B.4298 <u>O⁶-(4-chlorothenyl</u>)guanine	282	0.008	0.005	>16
B.4300 <u>O</u> ⁶ -(4-methoxythenyl)guanine	277	0.0165		0.83

TABLE 7 (continued)		3		(4) (4) (4) (4)
INACTIVATOR	M.Wt	150 (µм) hAT	кајј 150 (µМ)	Stabling 1.7.2 (ii) By Spec
B.4309 <u>O</u> ⁶ -(5-methylsulphonylthenyl)guanine	325	0.072		>16
B.4313 <u>Q</u> ⁶ -(5-bromo-3-thienylmethyl)guanine	326	0.0065	0.035	
B.4317 <u>O</u> ⁶ -(4-cyanothenyl)guanine	272	0.0028		>19
8,4318 <u>Q</u> ⁶ -(4,5-dichlorothenyl)guanine	348	0.015		2.5
8.4319 <u>0</u> ⁶ -(6-chloro-3-picolyl)guanine	277	0.2		>13
8.4320 <u>0</u> ⁶ -(5-bromo-3-picolyl)guanine	321	0.25		>13
B.4321 <u>O⁶</u> -(2-chloro-4-picolyl)guanine	277	0.04		>16

TABLE Z (continued)		(Atr.)		C+ability T 1/2 (b)
INACTIVATOR	Z.E.	150 (Jam) hAT	кај! 150 (µМ)	By Spec
8.4336 <u>Q⁶-(5-bromofurfury</u>])guanine	310	0.02		0.32
B.4354 <u>O</u> ⁶ -(4-isothiazolylmethyl)guanine	248	0.07	·	
B.4356 <u>Q⁶-(4-methy</u> lthiothenyl)guanine	293	0.0095		
B.4357 <u>Q</u> ⁶ -(5-iodo-3-thienylmethyl)guanine	447	0.009		>16
B.4361 <u>O</u> ⁶ -(4-methylsulphonylthenyl)guanine	325	0.2		>16
8.4366 0^6 -(naphtho[2,1- b]thiophen-2-ylmethyl)guanine	347	0.05		

IABLE 7 (continued)		(##)	ייים בין	Stability T 1/2 (h)
INACTIVATOR	₩. ₩.	+50 \hr.	05° (мп)	By Spec
B.4368 9-(8-D-arabinofuranosyl)- <u>Q</u> ⁶ -(4-bromothenyl)guanine	458	0.115		
B.4369 $\overline{0}^6$ -(4-bromothenyl)-9-(ethoxymethyl)guanine	384	0.28		
B.4370 $\underline{0}^6$ -(4-bromothenyl)-9(octyloxymethyl)guanine	468	1.2		
B.4373 <u>O</u> ⁶ -(4-azidothenyl)guanine	288	0.0063		
8.4377 <u>Q</u> ⁶ -(4-methylsulphinylthenyl)guanine	309	0.15		
B.4378 <u>O</u> ⁶ (5-phenylthenyl)guanine	323	0.75		
B.4379 <u>O</u> ⁶ -(4-bromothenyl)-2-deoxyguanosine	442	0.095		·

NACTIVATOR	M.Wt		In vitr	In vitro I ₅₀ (μM)	=	Raji I50	Stabili	Stability T 1/2 (h)
		hAT	тАТ	rAT	chAT	ogt	By Spec	By Assay
.4363 ⁵ -(4-bromothenyl)guanosine	458	0.08		0.24	0.95	30	>16	× × × × × × × × × × × × × × × × × × ×
lank space = not done								- 72

TABLE 8

ATASE ACTIVITY IN VARIOUS TISSUES OF NU/NU MICE AFTER TREATMENT WITH 10mg/kg (IP) B.4280
MEAN ACTIVITY (fm/mg)

	יייין אכן דודון אוואלן		
Tissue	<u>24h</u>	48h	Control*
Tumour	36 ± 7.79	140 ± 43.87	125
Liver	89.7 ± 10.14	100.7 ± 8.73	110**
Lung	15.3 ± 2.05	24 ± 2.83	43
Kidney	24.3 ± 4.03	28.7 ± 4.11	33
Spleen	41 ± 5.35	68.3 ± 9.53	. 81
Brain	13.7 ± 2.05	16.3 ± 1.25	14
Testis	45 ± 7.48	44 ± 1.41	45
Bone Marrow (pooled)	42	61	30

control values taken from a separate experiment** mean of 2 control liver values

Table 8. Effect of B.4280 on ATase activity in several tissues of nude mice. Animals were given a single dose of B.4280 (10mg/kg i.p.) and sacrificed 24 or 48 hours later.

TOXICITY OF INACTIVATORS IN COMBINATION WITH BCNU IN \mathtt{DBA}_2 MICE

	I			
	12mg/kg BCNU	50 (3/6)**	100 (6/6)**	100 (15/15)
SSURVIVAL AFTER 14 DAYS	16mg/kg BCNU	*(9/0) 0	50 (3/6)*	100 (15/15)
%SURVI	20mg/kg BCNU	33 (2/6)	(9/0) 0	93 (14/15)
INACTIVATOR	(60mg/kg)	06-benzylguanine	8.4205	8.4280

* 15mg/kg BCNU

All agents were given as a single i.p. dose

Table 9 Effect of ATase inactivators on the acute toxicity of bis-chloroethylnitrosourea (BCNU) in DBA2 mice.

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CLAIMS

A 6-hetarylalkyloxy pyrimidine derivative of formula II

5

10

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wherein

R is (i) a cyclic group having at least one 5- or 6-membered heterocyclic ring, optionally with a carbocyclic or heterocyclic ring fused thereto, the or each heterocyclic ring having at least one hetero atom chosen from 0, N, or S, or a substituted derivative thereof; or (iii) phenyl or a substituted derivative thereof,

R² is selected from H, C₁-C₅ alkyl, halogen or NH₂,
R⁴ and R⁵ which are the same or different are selected
from H, NH-Y' or NO_n
wherein
Y' is H, ribosyl, deoxyribosyl, arabinosyl, R''XCHR''' wherein
X is O or S, R'' is alkyl and R''' is H or alkyl, or
substituted derivatives thereof,

n = 1 or 2 or R^4 and R^5 together with the pyrimidine ring form a 5 or 6-membered ring structure containing one or more hetero atoms, and pharmaceutically acceptable salts thereof,

30 with the proviso that $\rm R^2$ is not NH $_2$ if $\rm R^4$ and $\rm R^5$ form a ring structure IX



wherein Y is H, ribosyl, deoxyribosyl, or R''XCHR''' wherein X is O or S, R'' and R''' are alkyl, or subsituted derivatives thereof,

5

and with the proviso that R is not phenyl in the following circumstances a) to h):

a) if R^2 and R^5 are NH_2 and R^4 is NO or NO_2

10

b) if R^2 is NH_2 and R^4 and R^5 form a ring structure X

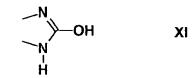
N N N

X

15

c) if R^2 is NH₂ and R^4 and R^5 form a ring structure XI

20



- d) if R^2 is NH_2 , R^4 is NO_2 and R^5 is H or CH_3
 - e) if R^2 , R^4 and R^5 are NH_2 .
 - f) if R^2 and R^5 are NH_2 and R^4 is H

30

- g) if R^2 is H, R^4 is NO_2 and R^5 is NH_2
- h) if R^2 is F or OH, and R^4 and R^5 form a ring structure XII

35



XII

2. A compound according to claim 1 which is of Formula III

10 wherein:

R is as defined in claim 1

Y' is as defined in claim 1; $R^2 \text{ is H, NH}_2, C_1 - C_5 \text{ alkyl or halogen;}$ 15 $R^3 \text{ is H or OH;}$

3. A compound according to claim 1 which is of Formula IV

20 RCH₂O X A IV

wherein:

30

25 R and Y' are as defined in claim 1;
X is CH or N;
A is CH or N;

4. A compound according to claim 1 which is of Formula ${f V}$

RCH₂O H N N N V wherein:

R is as defined in claim 1

X is CH or N

5 A is CH or N.

5. A compound according to claim 1 which is of Formula VI

10

wherein:

R is as defined in claim 1;

15

Z is 0 or S or CH = CH.

6. A compound according to claim 1 which is of Formula VII

20

wherein:

25 R is as defined in claim 1;

U is CH or N;

V is CH or N;

W is CH or N;

- 30 provided that U, V and W are not all CH.
 - 7. A compound according to claim 1 which is of Formula VIII

25

wherein:

R is as defined in claim 1 T is H, NH_2 or NO_n where n = 1 or 2; Q is H, NH_2 or NO_n where n = 1 or 2.

- 8. A compound according to claim 1, 2 or 3 wherein Y' is H, ribosyl, deoxyribosyl, arabinosyl, $HOCH_2CH_2OCH_2$ or R''XCHR''' wherein X is O or S, R'' and R''' are alkyl, or substituted derivatives thereof.
- 9. A compound according to claim 1 wherein R is a 5-membered heterocyclic ring, having at least one S atom therein.
- 15 10. A compound according to claim 1 wherein R is selected from a thiophene ring, a furan ring, and substituted derivatives thereof.
 - 11. A compound according to claim 1 wherein R includes a heterocyclic and/or carbocyclic ring substituted by halo, haloalkyl, cyano, SO_nR^8 where R^7 is alkyl and n = 0, 1 or 2, or $-COOR^8$ wherein R^8 is alkyl.
 - 12. A compound according to claim 1 wherein R is selected from a thiophene ring, a furan ring and substituted derivatives thereof selected from bromo- and cyano-substituted derivatives thereof.
 - 13. A compound according to claim 1 wherein R is selected from thiophene and furan rings with a chloro-, bromo- or cyano-substituent in a 1,3- or 1,4-relationship with the methyleneoxy group attached to the pyrimidine residue.
 - 14. A compound according to claim 1 wherein Y' is alkoxymethyl optionally substituted with OH on the alkyl of the alkoxy group.
- 35 15. A compound according to claim 1 which is selected from

 0^6 -(4-bromothenyl)-8-thiaguanine.

 0^4 -(4-chlorothenyl)pterin.

 0^6 -piperonyl -7-deaza-8-azaguanine.

 $5 \quad 0^6$ -(4-bromothenyl)-8-hydroxyguanine.

 0^6 -(4-chlorothenyl)-2, 4-diamino-6-hydroxy-5-nitrosopyrimidine.

16. Guanine derivatives of formula XIII

10

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wherein

15 E is 0 or S,

Y' is as defined in claim 1, R^6 is a cyclic group having at least one 5- or 6-membered heterocyclic ring, optionally with a carbocyclic or heterocyclic ring fused thereto, the or each heterocyclic ring having at least one hereto atom chosen from 0, N or S, or a substituted derivative thereof,

and pharmaceutically acceptable salts thereof, with the proviso that compounds published in WO 94/29312 are disclaimed.

- 25 17. A compound according to claim 16 wherein R^6 is a 5-membered heterocyclic ring, having at least one S atom therein.
 - 18. A compound according to claim 16 wherein \mathbf{R}^6 is selected from a thiophene ring, a furan ring, and substituted derivatives thereof.

30

19. A compound according to claim 16 wherein R^6 includes a heterocyclic and/or carbocyclic ring substituted by halo, haloalkyl, cyano, SO_nR^7 where R^7 is alkyl and n=0, 1 or 2, or $-COOR^8$ wherein R^8 is alkyl.

35

20. A compound according to claim 16 wherein \mathbf{R}^6 is selected from a thiophene ring, a furan ring and substituted derivatives thereof

selected from bromo- and cyano-substituted derivatives thereof.

- 21. A compound according to claim 16 wherein R⁶ is selected from thiophene and furan rings with a chloro-, bromo- or cyano-substituent in a 1,3- or 1,4-relationship with the methyleneoxy group attached to the pyrimidine residue.
 - 22. Guanine derivatives of formula XIV:

10

20

- wherein R^{10} is bromo, chloro or cyano, and Y' is as defined in claim 1.
 - 23. A compound according to claim 22 wherein Y' is H, ribosyl, deoxyribosyl, or R''XCHR''' wherein X is O or S, R'' is alkyl and R''' is H or alkyl, or substituted derivatives thereof.
 - 24. $\underline{0}^6$ -(4-bromothenyl)guanine.
- 25. A compound according to claim 22 which is 0^6 -(4-bromothenyl)-9-(2-hydroxyethoxymethyl)guanine.
 - 26. A compound according to claim 22 which is $9(B-D-arabinofuranosyl)-\underline{0}^{6}-(4-bromothenyl)$ guanine.
- 30 27. A compound according to claim 22 which is 0^6 -(4-bromothenyl)guanosine.
 - 28. A compound according to claim 22 which is $\underline{0}^6\text{-}(4\text{-bromothenyl})\text{-}2\text{-deoxyguanosine.}$

35

29. A compound according to claim 20 which is selected from:

15

- 0^6 -(5-chlorothenyl)guanine
- $\underline{0}^6$ -(5-cyanothenyl)guanine
- 0^6 -(5-methylsulphinylthenyl)guanine
 - 0^6 -(4-chlorothenyl)guanine
- 10 0^6 -(4-methoxythenyl)guanine
 - $\underline{0}^6$ -(5-bromo-3-thienylmethyl)guanine
 - 0^6 -(4-cyanothenyl)guanine
 - 0^6 -(4,5-dichlorothenyl)guanine
 - 30. A compound according to claim 20 which is selected from
- 20 $\underline{0}^6$ -(4-methylthiothenyl)guanine
 - 0^6 -(4-azidothenyl)guanine.
 - 31. A compound according to claim 16 which is selected from
- 25 $\underline{0}^6$ -(2-chloro-4-picolyl) quanine
 - 0^6 -(5-bromofurfuryl)guanine
- 30 32. A pharmaceutical composition comprising a compound according to any of claims 1 to 31 and a pharmaceutically acceptable excipient.
 - 33. A pharmaceutical composition according to claim 32 further comprising an alkylating agent.
 - 34. A composition according to claim 33 wherein the alkylating agent is selected from 1,3 bis (2-chloroethyl)-1-nitrosourea (BCNU)

and temozolomide.

35. A method for depleting 0^6 -alkylguanine-DNA alkyltransferase activity in a host comprising:

administering to the host an effective Q^6 -alkylguanine-DNA alkyltransferase activity depleting amount of a composition comprising a compound according to any of claims 1 to 31.

10

36. A method for treating tumour cells in a host comprising:

administering to the host a composition comprising an inactivator compound according to any of claims 1 to 31 in an amount effective to deplete Q^6 -alkylguanine -DNA alkyltransferase activity sufficiently to enhance the effectiveness of a chemotherapeutic akylating agent; and

administering to the host a composition comprising an alkylating agent in an amount which is cytotoxically effective in combination with the said inactivator compound.

- 37. Use of a compound according to any of claims 1 to 31 in the manufacture of a medicament for depleting $\underline{0}^6$ -alkylguanine-DNA alkyltransferase activity in tumour cells.
 - 38. A pharmaceutical composition comprising $\underline{0}^6$ -(4-bromothenyl)guanine and a pharmaceutically acceptable excipient.

- 39. A pharmaceutical composition according to claim 38 which is suitable for oral administration.
- 40. A pharmaceutical composition according to claim 38 further comprising an alkylating agent.
 - 41. A pharmaceutical composition according to claim 40 wherein the

akylating agent is selected from 1,3 bis(2-chloroethyl)-1-nitrosourea (BCNU) and temozolomide.

5 42. A method for depleting $\underline{0}^6$ -alkylguanine -DNA alkyltransferase activity in a host comprising:

administering to the host an effective Q^6 -alkylguanine-DNA alkyltransferase activity depleting amount of a composition comprising Q^6 -(4-bromothenyl)guanine.

43. A method for treating tumour cells in a host comprising:

administering to the host a composition comprising $\underline{0}^6 \text{ (4-bromothenyl)} \text{ guanine in an amount effective to deplete } \underline{0}^6 \text{-alkylguanine-DNA alkyltransferase activity sufficiently to enhance the effectiveness of a chemotherapeutic alkylating agent; and$

administering to the host a composition comprising an alkylating agent in an amount which is cytotoxically effective in combination with $\underline{0}^6$ -(4-bromothenyl)guanine.

44. Q^6 -(methylene[3 H]-(4-bromothenyl)guanine.

25

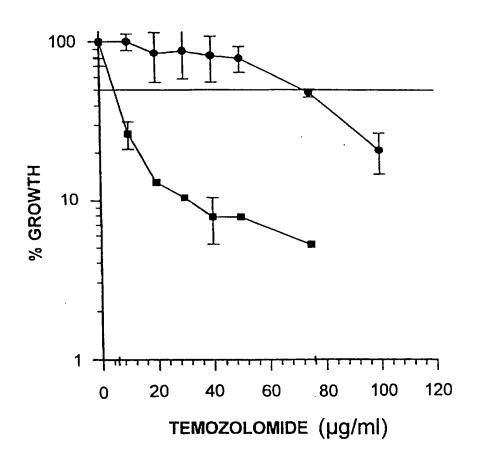
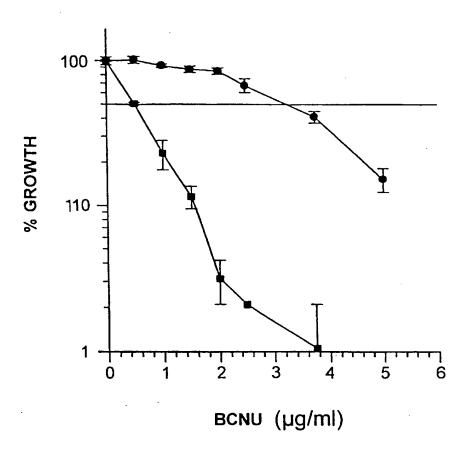




FIG. 1

WO 97/20843 PCT/IE96/00084

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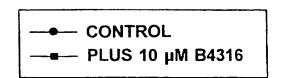


FIG. 2

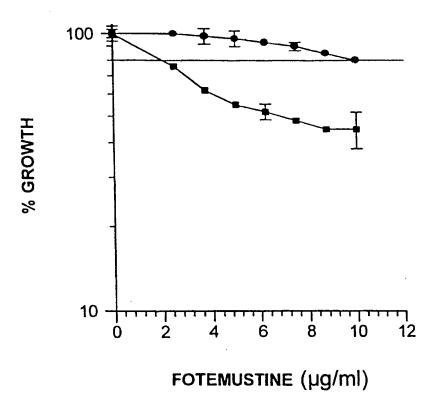




FIG. 3

WO 97/20843 PCT/IE96/00084

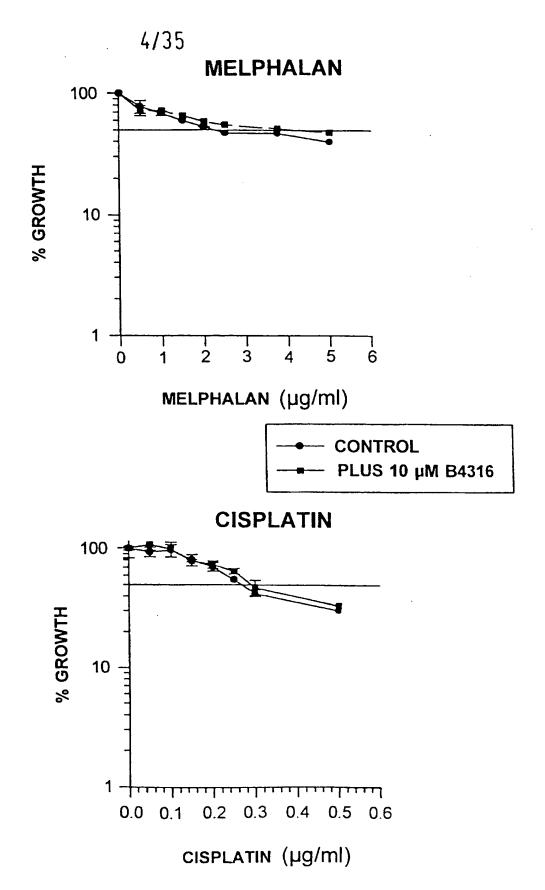
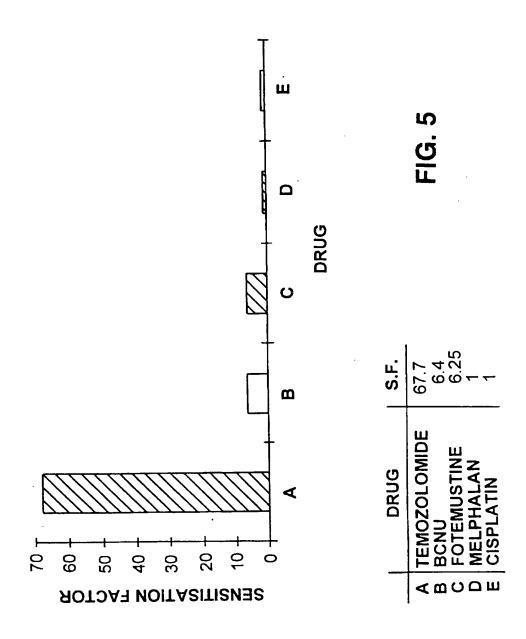
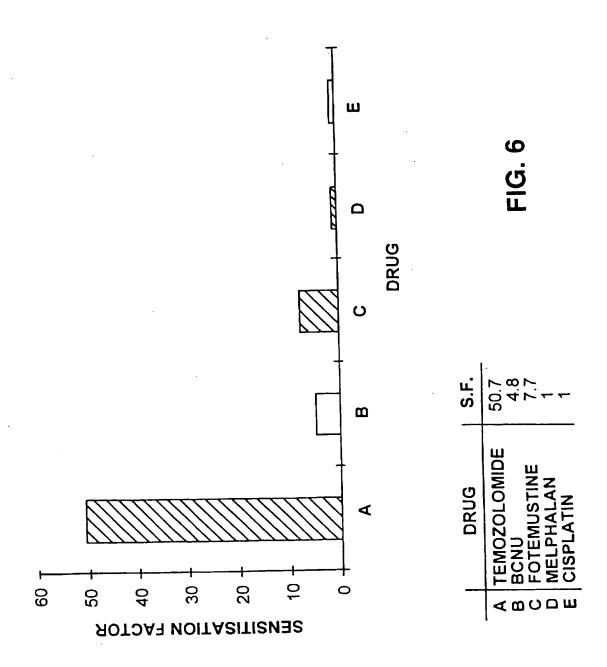


FIG. 4 SUBSTITUTE SHEET (RULE 26)





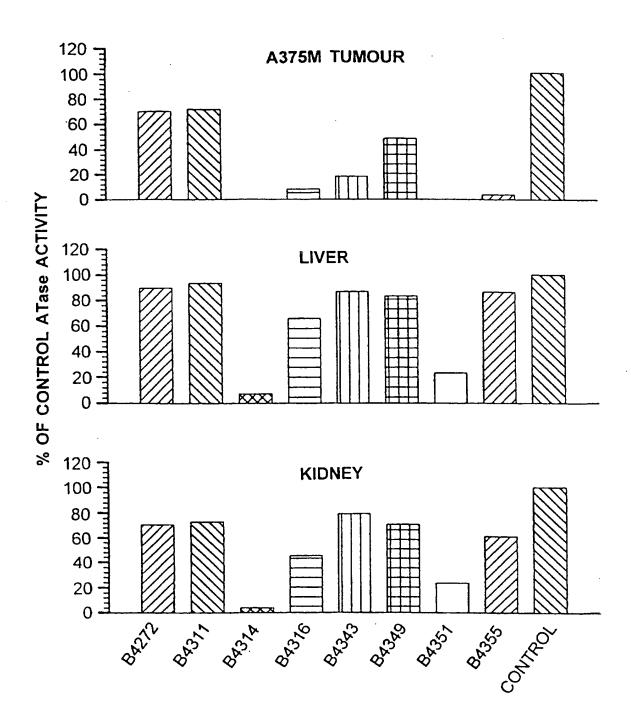
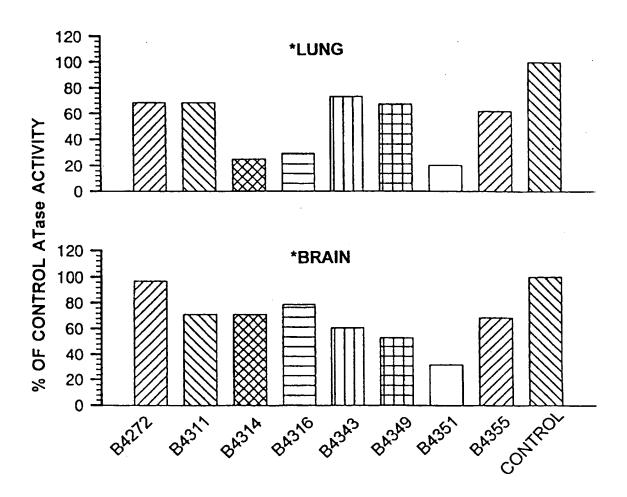


FIG. 7A



*HISTORICAL CONTROLS USED

FIG. 7B

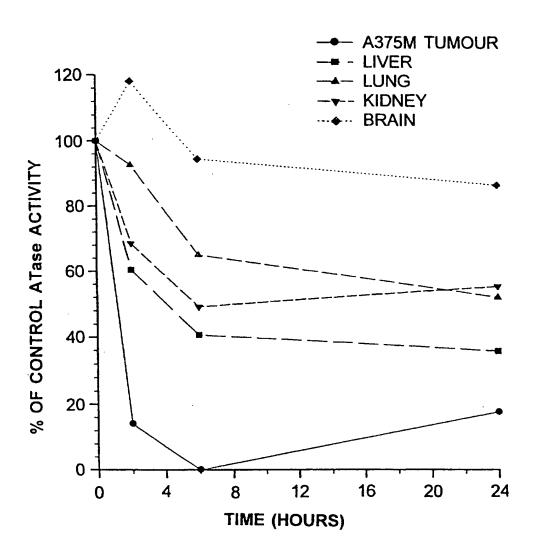


FIG. 8

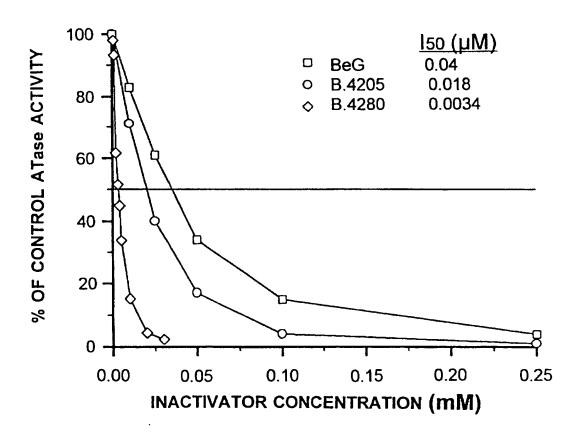
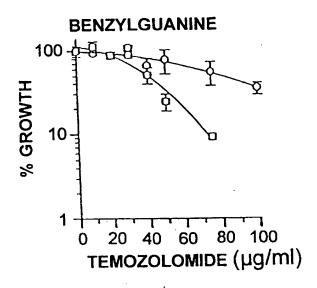
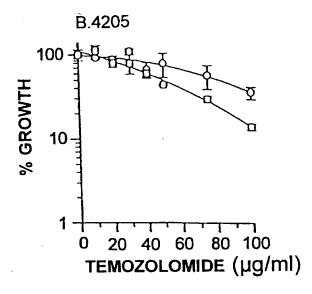


FIG. 9





□ 0.5µM INACTIVATOR



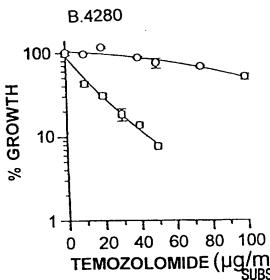
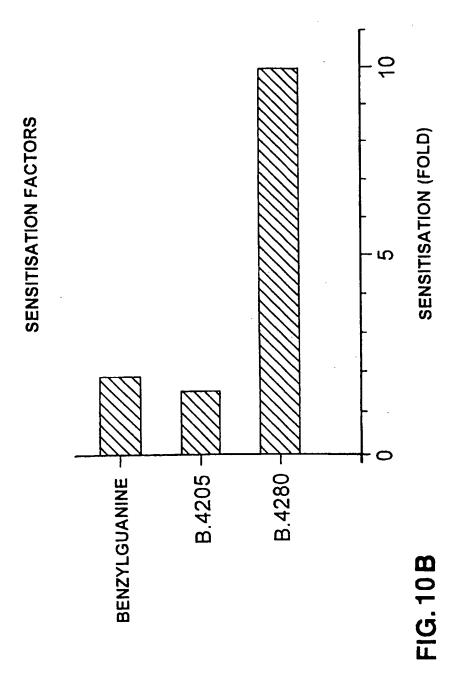
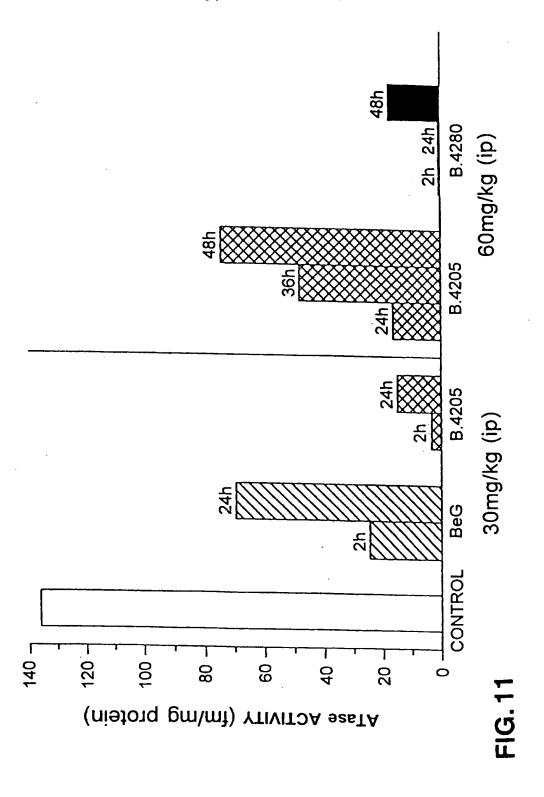
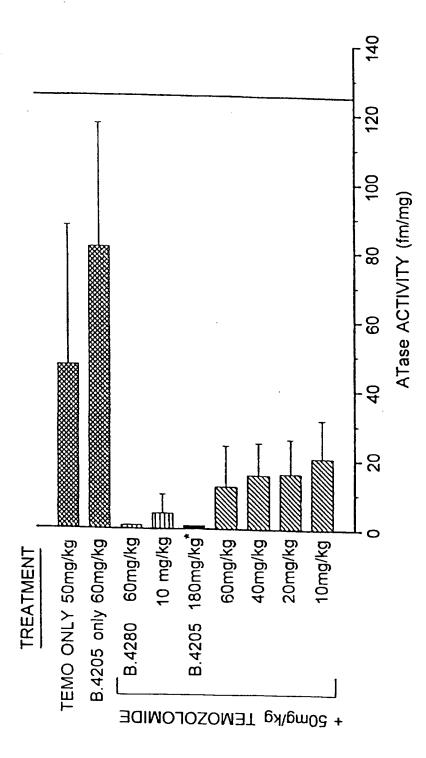


FIG. 10 A



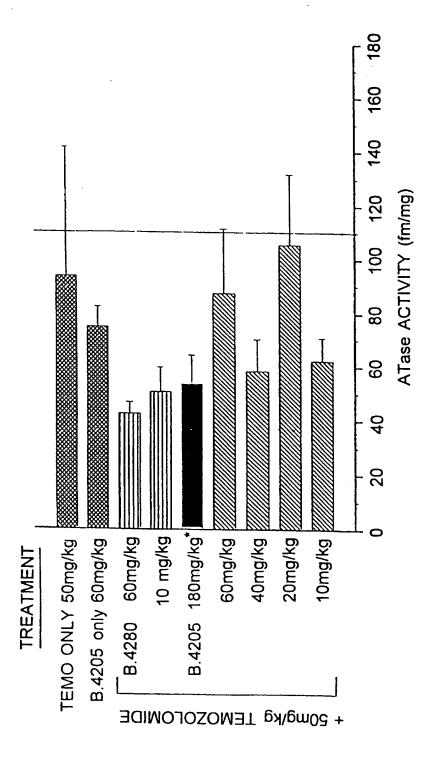
SUBSTITUTE SHEET (RULE 26)





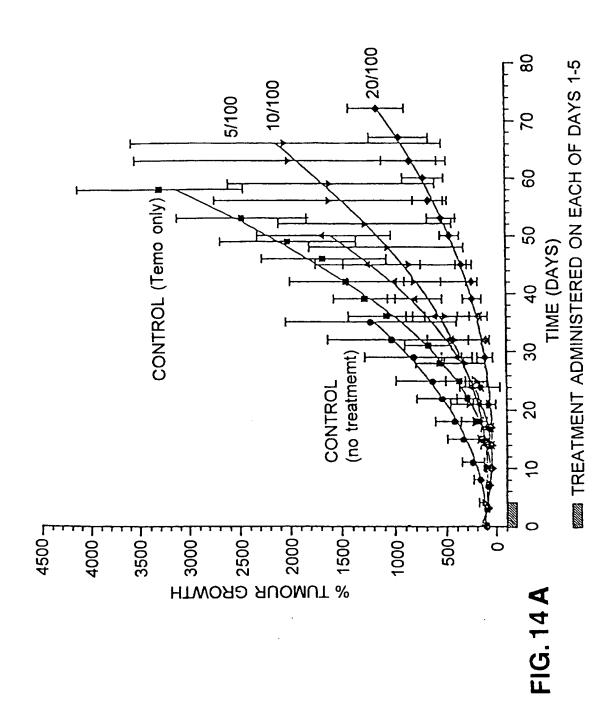
B.4205 + 50mg/kg TEMOZOLOMIDE GIVEN ON DAY 1 BY 50mg/kg TEMOZOLOMIDE ONLY ON DAYS 2 + 3 *180 mg/kg FOLLOWED E

FIG. 12

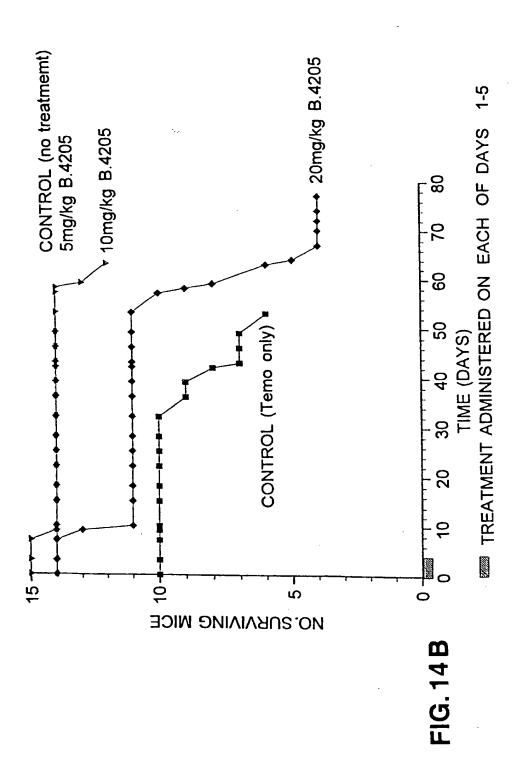


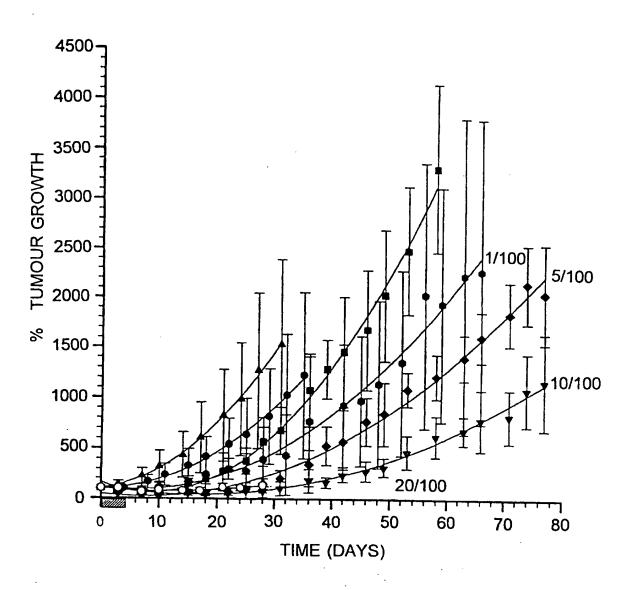
*180 mg/kg B.4205 + 50mg/kg TEMOZOLOMIDE GIVEN ON DAY 1 FOLLOWED BY 50mg/kg TEMOZOLOMIDE ONLY ON DAYS 2 + 3

FIG. 13



SUBSTITUTE SHEET (RULE 26)





TREATMENT ADMINISTERED ON THESE DAYS

- ▲ CONTROL (20mg/kg B4280)
- CONTROL (no treatment)
- CONTROL (Temo only)

FIG. 15A

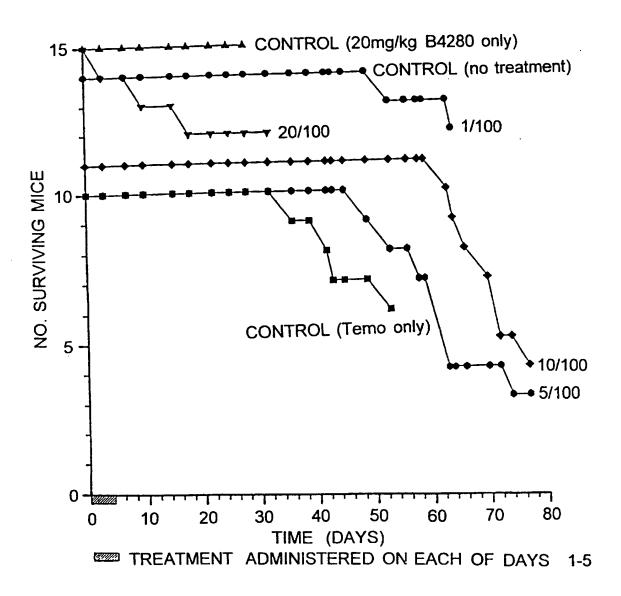
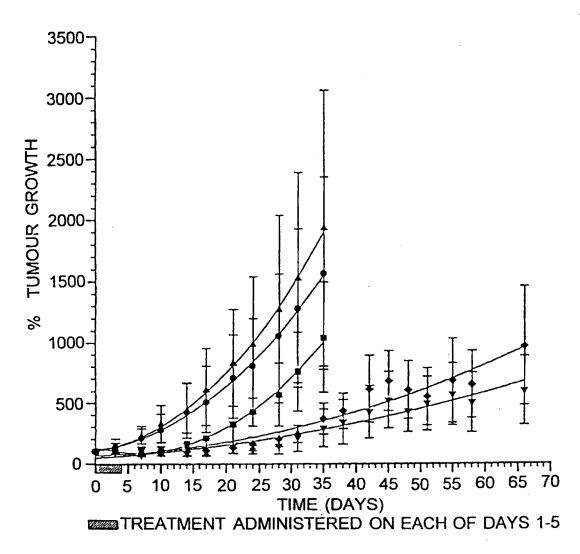
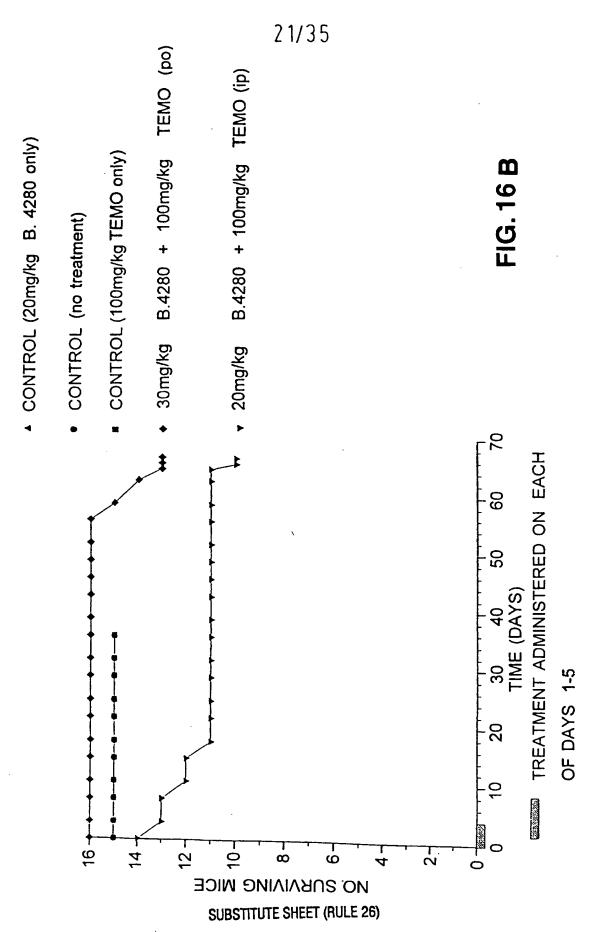


FIG. 15B

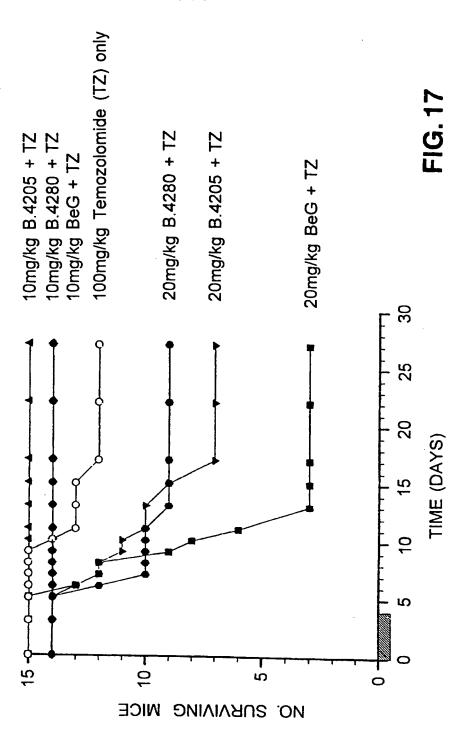


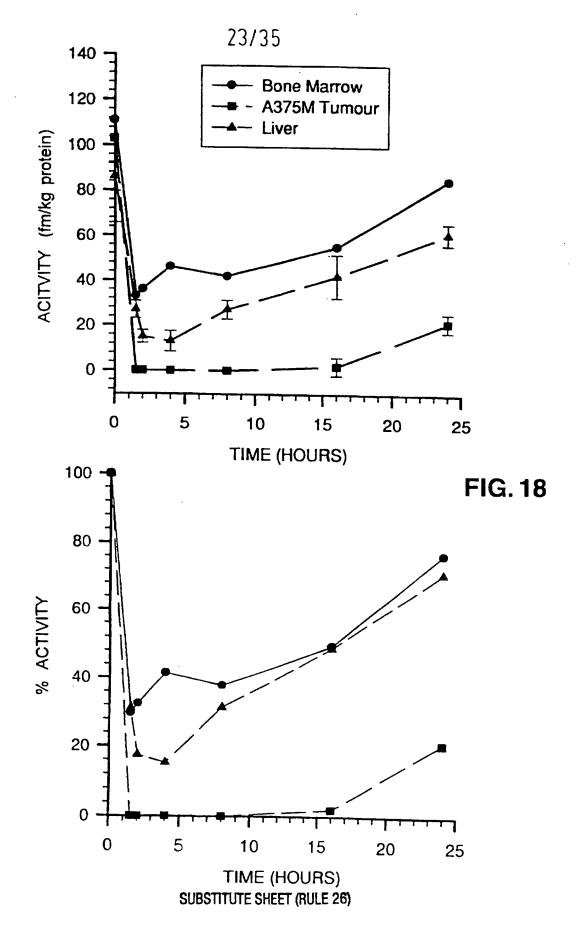
- ▲ CONTROL (20mg/kg B. 4280 only)
- CONTROL (no treatment)
- CONTROL (100mg/kg Temo only)
- + 30mg/kg B.4280 + 100mg/kg TEMO (po)
- ▼ 20mg/kg B.4280 + 100mg/kg TEMO (ip)

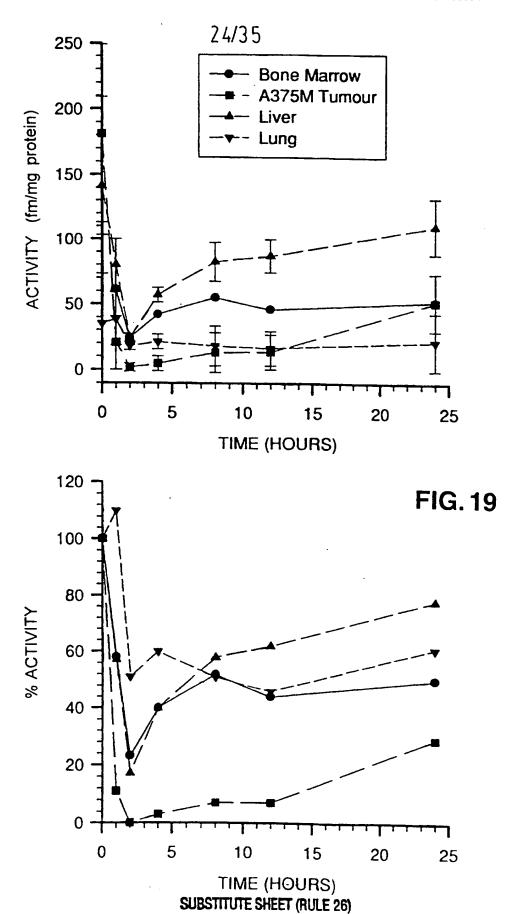
FIG. 16 A SUBSTITUTE SHEET (RULE 26)

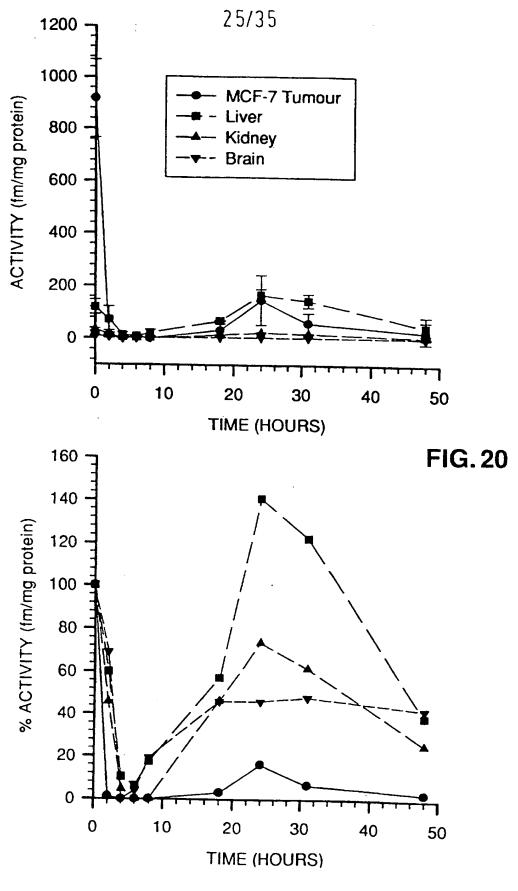




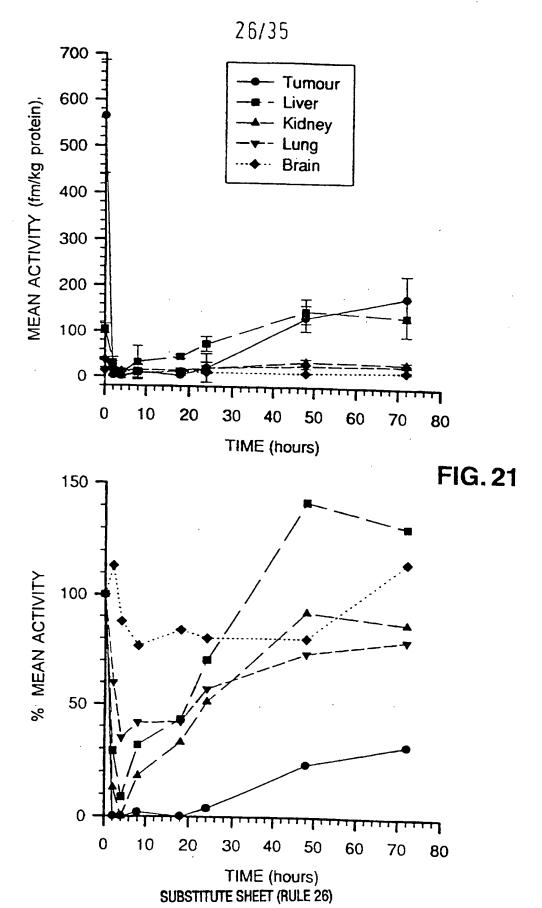


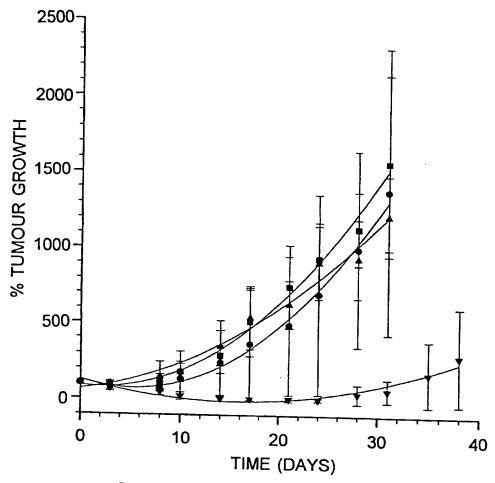






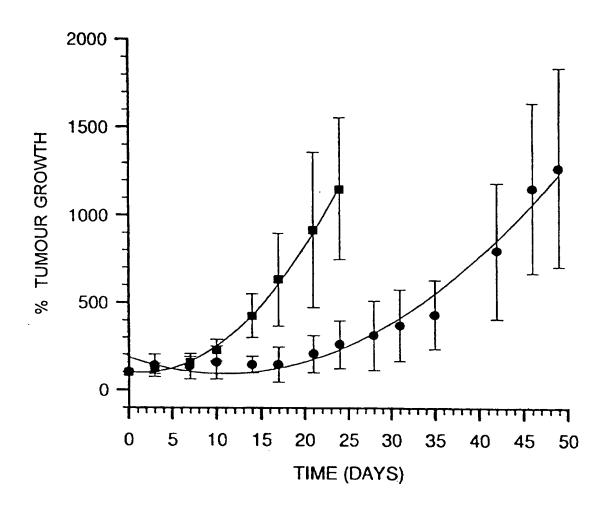
SUBSTITUTE SHEET (RULE 26)





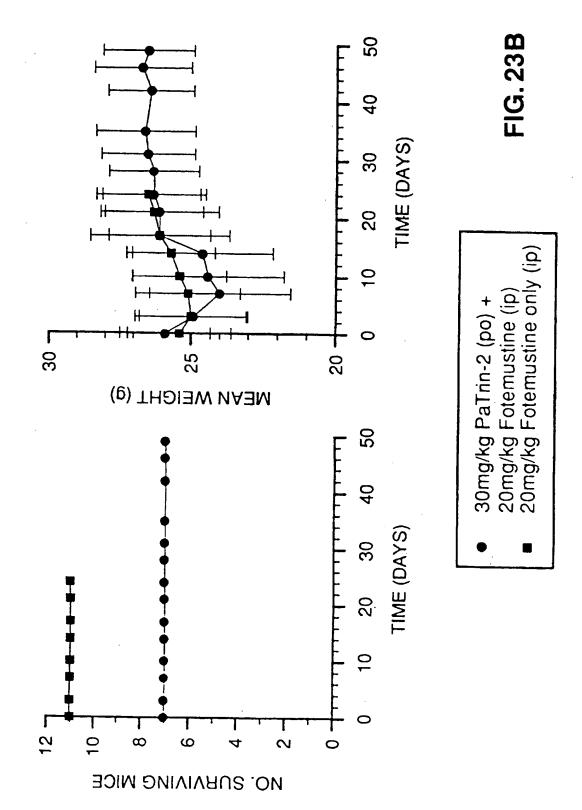
- Control (no treatment)
- 20mg/kg PaTrin-2 only
- ▲ 100mg/kg Temozolomide only
- ▼ 20mg/kg PaTrin-2 + 100mg/kg Temozolomide

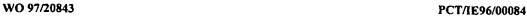
FIG. 22



- 30mg/kg PaTrin-2 (po) +
- 20mg/kg Fotemustine (ip)
 20mg/kg Fotemustine only (ip)

FIG. 23 A





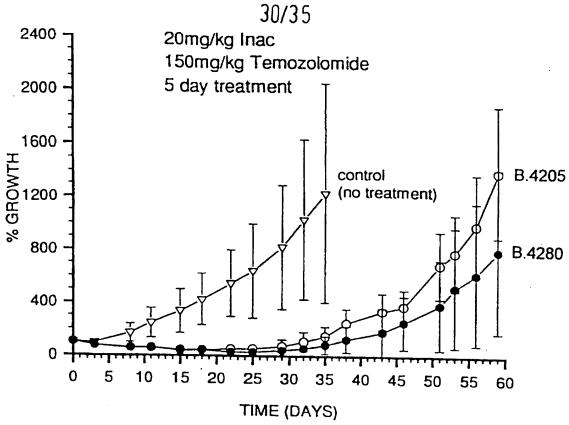
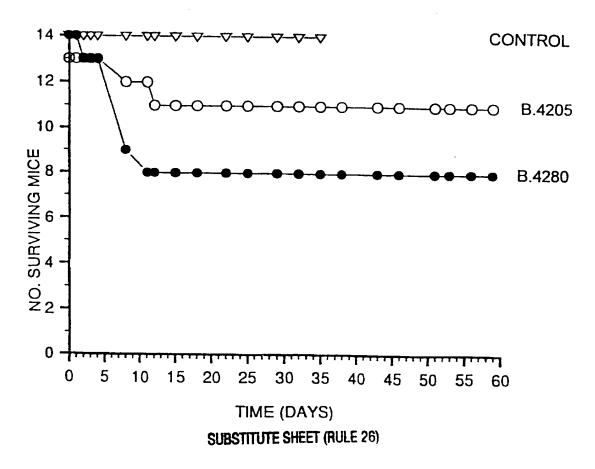
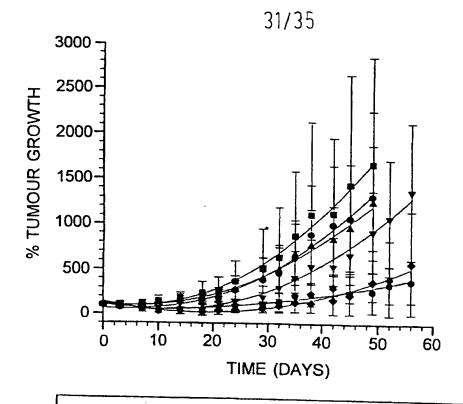
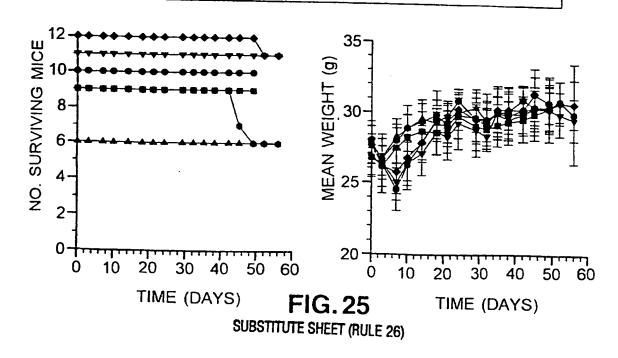


FIG. 24





- no treatment (v.c)
- 20mg/kg B4349 only
- ▲ 20mg/kg B4351 only
- ▼ 100mg/kg Temozolomide only
- 20mg/kg B4349 + 100mg/kg Temozolomide
- 20mg/kg B4351 + 100mg/kg Temozolomide



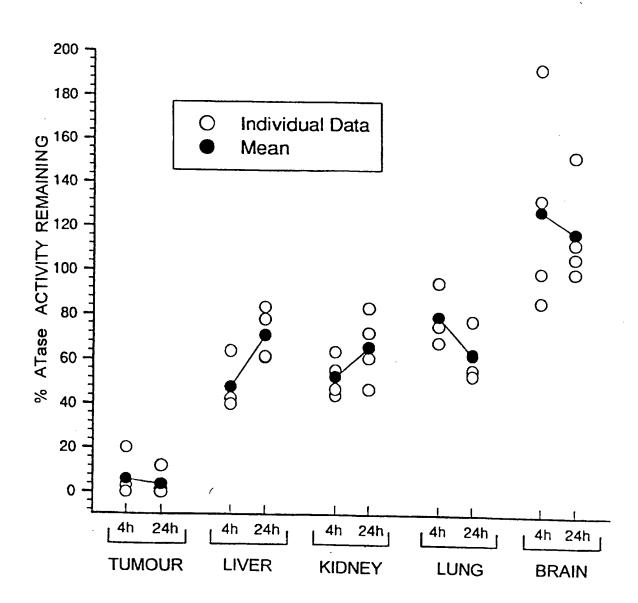


FIG. 26

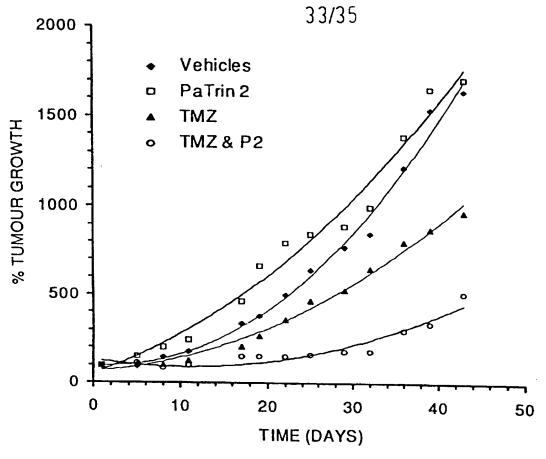
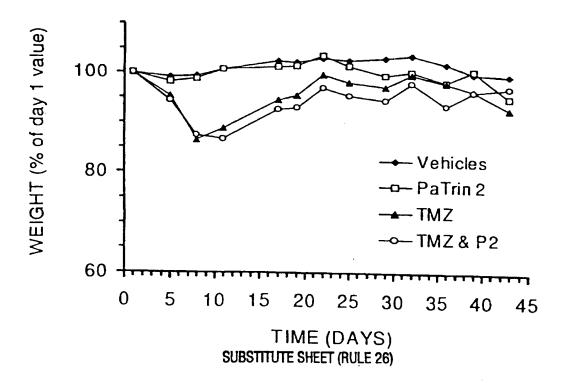


FIG. 27

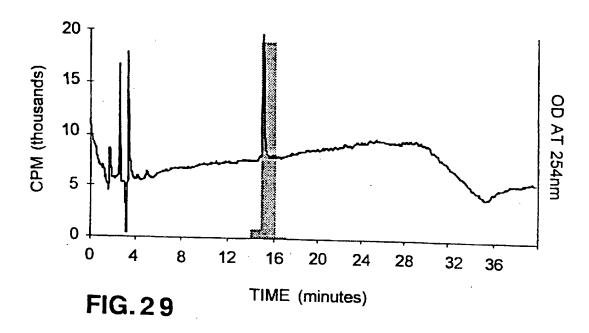


4-BROMO-2- THIOPHENECARBOXALDEHYDE

[3H]4-BROMOTHENYLALCOHOL

GUANINE SALT

[³H]4- BROMOTHENYLGUANINE SUBSTITUTE SHEET (RULE 26)



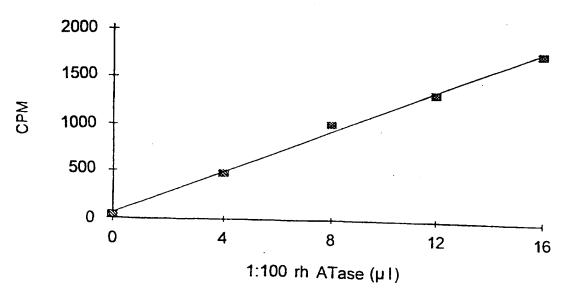


FIG. 30 SUBSTITUTE SHEET (RULE 26)

PCT/IE 96/00084

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IPC 6	FIGATION OF SUBJECT MATTER C07D473/18	7H19/16	C07D498/0	A61K31/505 C07D513/04
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C. DOCUM	I YTS CONSIDERED TO BE RELEVANT			
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c lateor	or other Apenal Peason (of Specified) and reforming to An oral dipploague, yan, exhibition or	CA	mot be considered	r relevance; the claimed invention to involve an inventive step when the I with one or more other such ducu-
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International Application No PCT/IE96/00084

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I sational application No.

PCT/IE 96/00084

Box 1 Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely: Although claims 35-36 are directed to a method of treatment of (diagnostic method practised on) the human/animal body the search has been carried out and based on the alleged effects of the compound/composition.
2. Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. As all searchable claims could be searches without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark on Protest The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

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Intc nal Application No PCT/IE 96/00084

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